

CXCR6 CHEMOKINE RECEPTOR GENE DISRUPTIONS, COMPOSITIONS AND METHODS RELATED THERETO

Related Applications

5 This application claims priority to U.S. Provisional Application No. 60/391,185, filed June 24, 2002 the entire contents of which are incorporated herein by reference.

Field of the Invention

10 The present invention relates to transgenic animals, compositions and methods relating to the characterization of gene function.

Background of the Invention

 Many medically significant biological processes are mediated by proteins participating in signal transduction pathways that involve G-proteins and/or second messengers such as cAMP. The membrane protein gene superfamily of G-protein
15 coupled receptors (GPCRs) include a wide range of biologically active receptors, such as hormone, viral, growth factor and neuroreceptors. GPCRs have been characterized as having seven putative transmembrane domains (designated TM1, TM2, TM3, TM4, TM5, TM6, and TM7), which are believed to represent transmembrane α -helices connected by extracellular or cytoplasmic loops. Most G-protein coupled receptors have
20 single conserved cysteine residues in each of the first two extracellular loops which form disulfide bonds that are believed to stabilize functional protein structure. G-protein coupled receptors can be intracellularly coupled by heterotrimeric G-proteins to various intracellular enzymes, ion channels and transporters. Different G-protein α -subunits preferentially stimulate particular effectors to modulate various biological functions in a
25 cell.

 Chemokines are chemotactic cytokines that are released by a wide variety of cells to attract macrophages, T cells, eosinophils, basophils and neutrophils to sites of inflammation (reviewed in Schall, Cytokine, 3, 165-183 (1991) and Murphy, Rev. Immun., 12, 593-633 (1994)). There are two classes of chemokines, C-X-C (α) and C-C
30 (β), depending on whether the first two cysteines are separated by a single amino acid (C-

X-C) or are adjacent (C-C). The α -chemokines are chemotactic primarily for neutrophils, whereas β -chemokines are chemotactic for macrophages, T-cells, eosinophils and basophils (Deng, *et al.*, *Nature*, 381, 661-666 (1996)).

The chemokines bind specific cell-surface receptors belonging to the family of G-protein-coupled seven-transmembrane-domain proteins (reviewed in Horuk, *Trends Pharm. Sci.*, 15, 159-165 (1994)), which are termed "chemokine receptors." Binding of a chemokine to its receptor induces a cascade of events, including generation of inositol triphosphate (IP₃), release of intracellular calcium, and activation of protein kinase C (PKC), resulting in changes in cell shape and changes in cell adhesiveness. These changes facilitate adhesion of the cell to endothelial cells (EC) lining blood vessel walls, which is followed by leukocyte transmigration between the EC into the tissues, where the leukocytes migrate along a gradient of increasing concentration of the chemokine to the site of release. Higher chemokine concentration at the site of inflammation causes more intracellular changes resulting in respiratory burst (generation of oxygen radicals) and release of granule contents.

One member of the chemokines receptor subfamily of GPCRs is chemokine (C-X-C) receptor 6 (CXCR6). CXCR6 is also referred to as BONZO, STRL33, and TYMSTR.

By RT-PCR on tumor-infiltrating lymphocyte lines with degenerate primers based on sequences of known chemokine receptors, Liao *et al.*, *J. Exp. Med.* 185: 2015-2023 (1997) isolated a cDNA encoding a GPCR, which they designated STRL33. Using an expression cloning strategy with a T-cell cDNA library, Deng *et al.*, *Nature* 388: 296-300 (1997) isolated an identical cDNA that they named BONZO. The predicted 342-amino acid STRL33/BONZO protein shares 25 to 30% amino acid identity with other chemokine receptors and is most similar to other GPCRs in the transmembrane domains. Northern blot analysis detected a 2.1-kb transcript in spleen, thymus, small intestine, and to a lesser extent in peripheral blood leukocytes, prostate, and colon; a 2.6-kb transcript was detected in placenta (Deng *et al.*, 1997). By fluorometric calcium flux analysis, Liao *et al.* (1997) were unable to detect responses in STRL33-expressing cells to a variety of chemokines. However, coexpression of CD4 and STRL33/BONZO rendered cells able to fuse with macrophage-tropic HIV-1 (transmission-type) strains as well as with simian

immunodeficiency virus. Liao et al. (1997) but not Deng et al. (1997) also detected fusion with T-cell-tropic HIV-1 strains. Liao et al. (1997) and Deng et al. (1997) speculated that expression of STRL33/BONZO may substitute for other HIV coreceptors, for example in CCR5 -/- individuals who become infected with HIV.

5 By expression cloning of mouse CXCL16, Matloubian et al., *Nature Immun.* 1: 298-304 (2000) identified a protein with 71% amino acid identity to human BONZO, which they renamed CXCR6. Human and mouse cells expressing CXCR6 showed a strong chemotactic response to CXCL16 but not to other chemokines. The authors concluded that CXCL16 and CXCR6 probably function in interactions between dendritic
10 cells and T cells and in regulating T-cell migration in the splenic red pulp. CXCL16 was also found in the thymic medulla and in some nonlymphoid tissues, indicating roles in thymocyte development and effector T cell trafficking.

By Southern blot analysis of genomic DNA and somatic cell hybrid analysis, Liao et al. (1997) mapped the single-copy STRL33 gene to chromosome 3. They suggested
15 that the STRL33 gene may be localized within the cluster of chemokine receptor-encoding genes in 3p21.

The complete 1251 bp cds nucleotide sequence for the murine CXCR6 gene has been deposited in GenBank (Accession: AF301018; GI: 11342661).

Given the importance of GPCRs, particularly chemokine receptors such as
20 CXCR6, a clear need exists for the elucidation of their functions, which information can be used in preventing, ameliorating or correcting dysfunctions or diseases associated therewith.

Summary of the Invention

Before the present compositions, antibodies, assays and methods for producing or
25 treating are disclosed and described, it is to be understood that this invention is not limited to particular compositions, antibodies, assays and methods for producing or treating and as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by
30 the appended claims.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

The present invention generally relates to transgenic animals, as well as to compositions and methods relating to the characterization of gene function.

The present invention provides transgenic cells comprising a disruption in a CXCR6 gene. The transgenic cells of the present invention are comprised of any cells capable of undergoing homologous recombination. Preferably, the cells of the present invention are stem cells and more preferably, embryonic stem (ES) cells, and most preferably, murine ES cells. According to one embodiment, the transgenic cells are produced by introducing a targeting construct into a stem cell to produce a homologous recombinant, resulting in a mutation of the CXCR6 gene. In another embodiment, the transgenic cells are derived from the transgenic animals described below. The cells derived from the transgenic animals includes cells that are isolated or present in a tissue or organ, and any cell lines or any progeny thereof.

The present invention also provides a targeting construct and methods of producing the targeting construct that when introduced into stem cells produces a homologous recombinant. In one embodiment, the targeting construct of the present invention comprises first and second polynucleotide sequences that are homologous to the CXCR6 gene. The targeting construct may also comprise a polynucleotide sequence that encodes a selectable marker that is preferably positioned between the two different homologous polynucleotide sequences in the construct. The targeting construct may also comprise other regulatory elements that can enhance homologous recombination.

The present invention further provides non-human transgenic animals and methods of producing such non-human transgenic animals comprising a disruption in a CXCR6 gene. The transgenic animals of the present invention include transgenic animals that are heterozygous and homozygous for a null mutation in the CXCR6 gene.

In one aspect, the transgenic animals of the present invention are defective in the function of the CXCR6 gene. In another aspect, the transgenic animals of the present invention comprise a phenotype associated with having a mutation in a CXCR6 gene. Preferably, the transgenic animals are rodents and, most preferably, are mice.

5 In a preferred embodiment, the present invention provides a transgenic mouse comprising a disruption in a CXCR6 gene, wherein there is no native expression of the endogenous CXCR6 gene.

 The present invention also provides methods of identifying agents capable of affecting a phenotype of a transgenic animal. For example, a putative agent is
10 administered to the transgenic animal and a response of the transgenic animal to the putative agent is measured and compared to the response of a “normal” or wild-type mouse, or alternatively compared to a transgenic animal control (without agent administration). The invention further provides agents identified according to such methods. The present invention also provides methods of identifying agents useful as
15 therapeutic agents for treating conditions associated with a disruption or other mutation (including naturally occurring mutations) of the CXCR6 gene.

 The present invention further provides a method of identifying agents having an effect on CXCR6 expression or function. The method includes administering an effective amount of the agent to a transgenic animal, preferably a mouse. The method
20 includes measuring a response of the transgenic animal, for example, to the agent, and comparing the response of the transgenic animal to a control animal, which may be, for example, a wild-type animal or alternatively, a transgenic animal control. Compounds that may have an effect on CXCR6 expression or function may also be screened against cells in cell-based assays, for example, to identify such compounds.

25 The invention also provides cell lines comprising nucleic acid sequences of a CXCR6 gene. Such cell lines may be capable of expressing such sequences by virtue of operable linkage to a promoter functional in the cell line. Preferably, expression of the CXCR6 gene sequence is under the control of an inducible promoter. Also provided are methods of identifying agents that interact with the CXCR6 gene, comprising the steps of
30 contacting the CXCR6 gene with an agent and detecting an agent/CXCR6 gene complex.

Such complexes can be detected by, for example, measuring expression of an operably linked detectable marker.

The invention further provides methods of treating diseases or conditions associated with a disruption in a CXCR6 gene, and more particularly, to a disruption or other alteration in the expression or function of the CXCR6 gene. In a preferred embodiment, methods of the present invention involve treating diseases or conditions associated with a disruption or other alteration in the CXCR6 gene's expression or function, including administering to a subject in need, a therapeutic agent that affects CXCR6 expression or function. In accordance with this embodiment, the method comprises administration of a therapeutically effective amount of a natural, synthetic, semi-synthetic, or recombinant CXCR6 gene, CXCR6 gene products or fragments thereof as well as natural, synthetic, semi-synthetic or recombinant analogs.

The present invention also provides compositions comprising or derived from ligands or other molecules or compounds that bind to or interact with CXCR6, including agonists or antagonists of CXCR6. Such agonists or antagonists of CXCR6 include antibodies and antibody mimetics, as well as other molecules that can readily be identified by routine assays and experiments well known in the art.

The present invention further provides methods of treating diseases or conditions associated with disrupted targeted gene expression or function, wherein the methods comprise detecting and replacing through gene therapy mutated or otherwise defective or abnormal CXCR6 genes.

Definitions

The term "gene" refers to (a) a gene containing at least one of the DNA sequences disclosed herein; (b) any DNA sequence that encodes the amino acid sequence encoded by the DNA sequences disclosed herein and/or; (c) any DNA sequence that hybridizes to the complement of the coding sequences disclosed herein. Preferably, the term includes coding regions, and preferably includes all sequences necessary for normal gene expression including promoters, enhancers and other regulatory sequences.

The term "neurological abnormality" or "neurological disorder" includes, for example, cognitive disorders, neuronal disorders, excitotoxicity and/or disorders which comprise neuronal apoptosis and/or excitotoxicity. Such neurological disorders may be

derived from (i) acute, subacute, or chronic injury to the nervous system, including traumatic injury, chemical injury, vascular injury and deficits (such as the ischemia resulting from stroke), together with infectious/inflammatory and tumor-induced injury; (ii) aging of the nervous system including, for example, Alzheimer's disease; (iii) chronic neurodegenerative diseases of the nervous system, including, for example, Parkinson's disease, Huntington's chorea, amyotrophic lateral sclerosis and the like, as well as spinocerebellar degenerations; and (iv) chronic immunological diseases of the nervous system or affecting the nervous system, including multiple sclerosis.

The terms "polynucleotide" and "nucleic acid molecule" are used interchangeably to refer to polymeric forms of nucleotides of any length. The polynucleotides may contain deoxyribonucleotides, ribonucleotides and/or their analogs. Nucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The term "polynucleotide" includes single-, double-stranded and triple helical molecules. "Oligonucleotide" refers to polynucleotides of between 5 and about 100 nucleotides of single- or double-stranded DNA. Oligonucleotides are also known as oligomers or oligos and may be isolated from genes, or chemically synthesized by methods known in the art. A "primer" refers to an oligonucleotide, usually single-stranded, that provides a 3'-hydroxyl end for the initiation of enzyme-mediated nucleic acid synthesis. The following are non-limiting embodiments of polynucleotides: a gene or gene fragment, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes and primers. A nucleic acid molecule may also comprise modified nucleic acid molecules, such as methylated nucleic acid molecules and nucleic acid molecule analogs. Analogs of purines and pyrimidines are known in the art, and include, but are not limited to, aziridinycytosine, 4-acetylcytosine, 5-fluorouracil, 5-bromouracil, 5-carboxymethylaminomethyl-2-thiouracil, 5-carboxymethyl-aminomethyluracil, inosine, N6-isopentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, pseudouracil, 5-pentyluracil and 2,6-diaminopurine. The use of uracil as a substitute for thymine in a deoxyribonucleic acid is also considered an analogous form of pyrimidine.

A "fragment" of a polynucleotide is a polynucleotide comprised of at least 9 contiguous nucleotides, preferably at least 15 contiguous nucleotides and more preferably at least 45 nucleotides, of coding or non-coding sequences.

The term "gene targeting" refers to a type of homologous recombination that occurs when a fragment of genomic DNA is introduced into a mammalian cell and that fragment locates and recombines with endogenous homologous sequences.

The term "homologous recombination" refers to the exchange of DNA fragments between two DNA molecules or chromatids at the site of homologous nucleotide sequences.

The term "homologous" as used herein denotes a characteristic of a DNA sequence having at least about 70 percent sequence identity as compared to a reference sequence, typically at least about 85 percent sequence identity, preferably at least about 95 percent sequence identity, and more preferably about 98 percent sequence identity, and most preferably about 100 percent sequence identity as compared to a reference sequence. Homology can be determined using, for example, a "BLASTN" algorithm. It is understood that homologous sequences can accommodate insertions, deletions and substitutions in the nucleotide sequence. Thus, linear sequences of nucleotides can be essentially identical even if some of the nucleotide residues do not precisely correspond or align. The reference sequence may be a subset of a larger sequence, such as a portion of a gene or flanking sequence, or a repetitive portion of a chromosome.

The term "target gene" (alternatively referred to as "target gene sequence" or "targeting DNA" or "target sequence") refers to any nucleic acid molecule, polynucleotide, or gene to be modified by homologous recombination. The target sequence includes an intact gene, an exon or intron, a regulatory sequence or any region between genes. The target gene may comprise a portion of a particular gene or genetic locus in the individual's genomic DNA. As provided herein, the target gene of the present invention is preferably the endogenous CXCR6 gene.

"Disruption" of a CXCR6 gene occurs when a fragment of genomic DNA locates and recombines with an endogenous homologous sequence. These sequence disruptions or modifications may include insertions, missense, frameshift, deletion, or substitutions, or replacements of DNA sequence, or any combination thereof. Insertions include the

insertion of entire genes, which may be of animal, plant, fungal, insect, prokaryotic, or viral origin. Disruption, for example, can alter the normal gene product by inhibiting its production partially or completely or by enhancing the normal gene product's activity. In a preferred embodiment, the disruption is a null disruption, wherein there is no significant expression of the CXCR6 gene.

The term "native expression" refers to the expression of the full-length polypeptide encoded by the CXCR6 gene, at expression levels present in the wild-type mouse. Thus, a disruption in which there is "no native expression" of the endogenous CXCR6 gene refers to a partial or complete reduction of the expression of at least a portion of a polypeptide encoded by an endogenous CXCR6 gene of a single cell, selected cells, or all of the cells of a mammal. The term "knockout" is a synonym for functional inactivation of the gene.

The term "construct" or "targeting construct" refers to an artificially assembled DNA segment to be transferred into a target tissue, cell line or animal. Typically, the targeting construct will include a gene or a nucleic acid sequence of particular interest, a marker gene and appropriate control sequences. As provided herein, the targeting construct of the present invention comprises a CXCR6 targeting construct. A "CXCR6 targeting construct" includes a DNA sequence homologous to at least one portion of a CXCR6 gene and is capable of producing a disruption in a CXCR6 gene in a host cell.

The term "transgenic cell" refers to a cell containing within its genome a CXCR6 gene that has been disrupted, modified, altered, or replaced completely or partially by the method of gene targeting.

The term "transgenic animal" refers to an animal that contains within its genome a specific gene that has been disrupted or otherwise modified or mutated by the method of gene targeting. "Transgenic animal" includes both the heterozygous animal (*i.e.*, one defective allele and one wild-type allele) and the homozygous animal (*i.e.*, two defective alleles).

As used herein, the terms "selectable marker" and "positive selection marker" refer to a gene encoding a product that enables only the cells that carry the gene to survive and/or grow under certain conditions. For example, plant and animal cells that express the introduced neomycin resistance (*Neo^r*) gene are resistant to the compound

G418. Cells that do not carry the *Neo^r* gene marker are killed by G418. Other positive selection markers are known to, or are within the purview of, those of ordinary skill in the art.

A "host cell" includes an individual cell or cell culture that can be or has been a recipient for vector(s) or for incorporation of nucleic acid molecules and/or proteins. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent due to natural, accidental, or deliberate mutation. A host cell includes cells transfected with the constructs of the present invention.

The term "modulates" or "modulation" as used herein refers to the decrease, inhibition, reduction, amelioration, increase or enhancement of a CXCR6 function, expression, activity, or alternatively a phenotype associated with a disruption in a CXCR6 gene. The term "ameliorates" or "amelioration" as used herein refers to a decrease, reduction or elimination of a condition, disease, disorder, or phenotype, including an abnormality or symptom associated with a disruption in a CXCR6 gene.

The term "abnormality" refers to any disease, disorder, condition, or phenotype in which a disruption of a CXCR6 gene is implicated, including pathological conditions and behavioral observations.

The term "pain" refers to all types of pain, including somatic pain, e.g., visceral pain or cutaneous pain, or pain caused by a burn, a bruise, an abrasion, a laceration, a broken bone, a torn ligament, a torn tendon, a torn muscle, a viral infection, a bacterial infection, a protozoal infection, a fungal infection, contact dermatitis, inflammation, or cancer; and neuropathic pain, e.g. caused by injury to the central or peripheral nervous system due to cancer, HIV infection, tissue trauma, infection, autoimmune disease, diabetes, arthritis, diabetic neuropathy, trigeminal neuralgia or drug administration.

Brief Description of the Drawings

Figure 1 shows the polynucleotide sequence for a murine CXCR6 gene (SEQ ID NO:1).

Figure 2 shows the amino acid sequence for murine CXCR6 (SEQ ID NO:2).

Figures 3-4 show the location and extent of the disrupted portion of the CXCR6 gene, as well as the nucleotide sequences flanking the *Neo^r* insert in the targeting

construct. Figure 4 shows the sequences identified as SEQ ID NO:3 and SEQ ID NO:4, which were used as the 5'- and 3'- targeting arms (including the homologous sequences) in the CXCR6 targeting construct, respectively.

Detailed Description of the Invention

5 The invention is based, in part, on the evaluation of the expression and role of genes and gene expression products, primarily those associated with a CXCR6 gene. Among other uses or applications, the invention permits the definition of disease pathways and the identification of diagnostically and therapeutically useful targets. For example, genes that are mutated or down-regulated under disease conditions may be
10 involved in causing or exacerbating the disease condition. Treatments directed at up-regulating the activity of such genes or treatments that involve alternate pathways, may ameliorate the disease condition.

Generation of Targeting Construct

 The targeting construct of the present invention may be produced using standard
15 methods known in the art. (*see, e.g.,* Sambrook, *et al.*, 1989, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York; E.N. Glover (eds.), 1985, *DNA Cloning: A Practical Approach*, Volumes I and II; M.J. Gait (ed.), 1984, *Oligonucleotide Synthesis*; B.D. Hames & S.J. Higgins (eds.), 1985, *Nucleic Acid Hybridization*; B.D. Hames & S.J. Higgins (eds.),
20 1984, *Transcription and Translation*; R.I. Freshney (ed.), 1986, *Animal Cell Culture*; Immobilized Cells and Enzymes, IRL Press, 1986; B. Perbal, 1984, *A Practical Guide To Molecular Cloning*; F.M. Ausubel *et al.*, 1994, *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc.). For example, the targeting construct may be prepared in accordance with conventional ways, where sequences may be synthesized, isolated from
25 natural sources, manipulated, cloned, ligated, subjected to *in vitro* mutagenesis, primer repair, or the like. At various stages, the joined sequences may be cloned, and analyzed by restriction analysis, sequencing, or the like.

 The targeting DNA can be constructed using techniques well known in the art. For example, the targeting DNA may be produced by chemical synthesis of
30 oligonucleotides, nick-translation of a double-stranded DNA template, polymerase chain-reaction amplification of a sequence (or ligase chain reaction amplification), purification

of prokaryotic or target cloning vectors harboring a sequence of interest (*e.g.*, a cloned cDNA or genomic DNA, synthetic DNA or from any of the aforementioned combination) such as plasmids, phagemids, YACs, cosmids, bacteriophage DNA, other viral DNA or replication intermediates, or purified restriction fragments thereof, as well as other
5 sources of single and double-stranded polynucleotides having a desired nucleotide sequence. Moreover, the length of homology may be selected using known methods in the art. For example, selection may be based on the sequence composition and complexity of the predetermined endogenous target DNA sequence(s).

The targeting construct of the present invention typically comprises a first
10 sequence homologous to a portion or region of the CXCR6 gene and a second sequence homologous to a second portion or region of the CXCR6 gene. The targeting construct may further comprise a positive selection marker, which is preferably positioned in between the first and the second DNA sequences that are homologous to a portion or region of the target DNA sequence. The positive selection marker may be operatively
15 linked to a promoter and a polyadenylation signal.

Other regulatory sequences known in the art may be incorporated into the targeting construct to disrupt or control expression of a particular gene in a specific cell type. In addition, the targeting construct may also include a sequence coding for a screening marker, for example, green fluorescent protein (GFP), or another modified
20 fluorescent protein.

Although the size of the homologous sequence is not critical and can range from as few as about 15-20 base pairs to as many as 100 kb, preferably each fragment is greater than about 1 kb in length, more preferably between about 1 and about 10 kb, and even more preferably between about 1 and about 5 kb. One of skill in the art will
25 recognize that although larger fragments may increase the number of homologous recombination events in ES cells, larger fragments will also be more difficult to clone.

In a preferred embodiment of the present invention, the targeting construct is prepared directly from a plasmid genomic library using the methods described in pending U.S. Patent Application Ser. No.: 08/971,310, filed November 17, 1997, the disclosure
30 of which is incorporated herein in its entirety. Generally, a sequence of interest is identified and isolated from a plasmid library in a single step using, for example, long-

range PCR. Following isolation of this sequence, a second polynucleotide that will disrupt the target sequence can be readily inserted between two regions encoding the sequence of interest. In accordance with this aspect, the construct is generated in two steps by (1) amplifying (for example, using long-range PCR) sequences homologous to the target sequence, and (2) inserting another polynucleotide (for example a selectable marker) into the PCR product so that it is flanked by the homologous sequences. Typically, the vector is a plasmid from a plasmid genomic library. The completed construct is also typically a circular plasmid.

In another embodiment, the targeting construct is designed in accordance with the regulated positive selection method described in U.S. Patent Application Ser. No. 09/954,483, filed September 17, 2001, the disclosure of which is incorporated herein in its entirety. The targeting construct is designed to include a PGK-*neo* fusion gene having two *lacO* sites, positioned in the PGK promoter and an NLS-*lacI* gene comprising a lac repressor fused to sequences encoding the NLS from the SV40 T antigen.

In another embodiment, the targeting construct may contain more than one selectable maker gene, including a negative selectable marker, such as the herpes simplex virus tk (HSV-tk) gene. The negative selectable marker may be operatively linked to a promoter and a polyadenylation signal. (*see, e.g.*, U.S. Patent No. 5,464,764; U.S. Patent No. 5,487,992; U.S. Patent No. 5,627,059; and U.S. Patent No. 5,631,153).

Generation of Cells and Confirmation of Homologous Recombination Events

Once an appropriate targeting construct has been prepared, the targeting construct may be introduced into an appropriate host cell using any method known in the art.

Various techniques may be employed in the present invention, including, for example: pronuclear microinjection; retrovirus mediated gene transfer into germ lines; gene

targeting in embryonic stem cells; electroporation of embryos; sperm-mediated gene transfer; and calcium phosphate/DNA co-precipitates, microinjection of DNA into the nucleus, bacterial protoplast fusion with intact cells, transfection, polycations, *e.g.*, polybrene, polyornithine, *etc.*, or the like (*see, e.g.*, U.S. Patent No. 4,873,191; Van der Putten, *et al.*, 1985, *Proc. Natl. Acad. Sci., USA* 82:6148-6152; Thompson, *et al.*, 1989, *Cell* 56:313-321; Lo, 1983, *Mol Cell. Biol.* 3:1803-1814; Lavitrano, *et al.*, 1989, *Cell*, 57:717-723). Various techniques for transforming mammalian cells are known in the art.

(see, e.g., Gordon, 1989, *Intl. Rev. Cytol.*, 115:171-229; Keown *et al.*, 1989, *Methods in Enzymology*; Keown *et al.*, 1990, *Methods and Enzymology*, Vol. 185, pp. 527-537; Mansour *et al.*, 1988, *Nature*, 336:348-352).

In a preferred aspect of the present invention, the targeting construct is introduced into host cells by electroporation. In this process, electrical impulses of high field strength reversibly permeabilize biomembranes allowing the introduction of the construct. The pores created during electroporation permit the uptake of macromolecules such as DNA. (see, e.g., Potter, H., *et al.*, 1984, *Proc. Nat'l. Acad. Sci. U.S.A.* 81:7161-7165).

Any cell type capable of homologous recombination may be used in the practice of the present invention. Examples of such target cells include cells derived from vertebrates including mammals such as humans, bovine species, ovine species, murine species, simian species, and ether eucaryotic organisms such as filamentous fungi, and higher multicellular organisms such as plants.

Preferred cell types include embryonic stem (ES) cells, which are typically obtained from pre-implantation embryos cultured *in vitro*. (see, e.g., Evans, M. J., *et al.*, 1981, *Nature* 292:154-156; Bradley, M. O., *et al.*, 1984, *Nature* 309:255-258; Gossler *et al.*, 1986, *Proc. Natl. Acad. Sci. USA* 83:9065-9069; and Robertson, *et al.*, 1986, *Nature* 322:445-448). The ES cells are cultured and prepared for introduction of the targeting construct using methods well known to the skilled artisan. (see, e.g., Robertson, E. J. ed. "Teratocarcinomas and Embryonic Stem Cells, a Practical Approach", IRL Press, Washington D.C., 1987; Bradley *et al.*, 1986, *Current Topics in Devel. Biol.* 20:357-371; by Hogan *et al.*, in "Manipulating the Mouse Embryo": A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor N.Y., 1986; Thomas *et al.*, 1987, *Cell* 51:503; Koller *et al.*, 1991, *Proc. Natl. Acad. Sci. USA*, 88:10730; Dorin *et al.*, 1992, *Transgenic Res.* 1:101; and Veis *et al.*, 1993, *Cell* 75:229). The ES cells that will be inserted with the targeting construct are derived from an embryo or blastocyst of the same species as the developing embryo into which they are to be introduced. ES cells are typically selected for their ability to integrate into the inner cell mass and contribute to the germ line of an individual when introduced into the mammal in an embryo at the

blastocyst stage of development. Thus, any ES cell line having this capability is suitable for use in the practice of the present invention.

The present invention may also be used to knock out or otherwise modify or disrupt genes in other cell types, such as stem cells. By way of example, stem cells may be myeloid, lymphoid, or neural progenitor and precursor cells. These cells comprising a knock out, modification or disruption of a gene may be particularly useful in the study of CXCR6 gene function in individual developmental pathways. Stem cells may be derived from any vertebrate species, such as mouse, rat, dog, cat, pig, rabbit, human, non-human primates and the like.

After the targeting construct has been introduced into cells, the cells in which successful gene targeting has occurred are identified. Insertion of the targeting construct into the targeted gene is typically detected by identifying cells for expression of the marker gene. In a preferred embodiment, the cells transformed with the targeting construct of the present invention are subjected to treatment with an appropriate agent that selects against cells not expressing the selectable marker. Only those cells expressing the selectable marker gene survive and/or grow under certain conditions. For example, cells that express the introduced neomycin resistance gene are resistant to the compound G418, while cells that do not express the neo gene marker are killed by G418. If the targeting construct also comprises a screening marker such as GFP, homologous recombination can be identified through screening cell colonies under a fluorescent light. Cells that have undergone homologous recombination will have deleted the GFP gene and will not fluoresce.

If a regulated positive selection method is used in identifying homologous recombination events, the targeting construct is designed so that the expression of the selectable marker gene is regulated in a manner such that expression is inhibited following random integration but is permitted (derepressed) following homologous recombination. More particularly, the transfected cells are screened for expression of the *neo* gene, which requires that (1) the cell was successfully electroporated, and (2) *lac* repressor inhibition of *neo* transcription was relieved by homologous recombination. This method allows for the identification of transfected cells and homologous recombinants to occur in one step with the addition of a single drug.

Alternatively, a positive-negative selection technique may be used to select homologous recombinants. This technique involves a process in which a first drug is added to the cell population, for example, a neomycin-like drug to select for growth of transfected cells, *i.e.* positive selection. A second drug, such as FIAU is subsequently added to kill cells that express the negative selection marker, *i.e.* negative selection. Cells that contain and express the negative selection marker are killed by a selecting agent, whereas cells that do not contain and express the negative selection marker survive. For example, cells with non-homologous insertion of the construct express HSV thymidine kinase and therefore are sensitive to the herpes drugs such as gancyclovir (GANC) or FIAU (1-(2-deoxy 2-fluoro-B-D-arabinofluranosyl)-5-iodouracil). (*see, e.g., Mansour et al., Nature* 336:348-352: (1988); Capecchi, *Science* 244:1288-1292, (1989); Capecchi, *Trends in Genet.* 5:70-76 (1989)).

Successful recombination may be identified by analyzing the DNA of the selected cells to confirm homologous recombination. Various techniques known in the art, such as PCR and/or Southern analysis may be used to confirm homologous recombination events.

Homologous recombination may also be used to disrupt genes in stem cells, and other cell types, which are not totipotent embryonic stem cells. By way of example, stem cells may be myeloid, lymphoid, or neural progenitor and precursor cells. Such transgenic cells may be particularly useful in the study of CXCR6 gene function in individual developmental pathways. Stem cells may be derived from any vertebrate species, such as mouse, rat, dog, cat, pig, rabbit, human, non-human primates and the like.

In cells that are not totipotent, it may be desirable to knock out both copies of the target using methods that are known in the art. For example, cells comprising homologous recombination at a target locus that have been selected for expression of a positive selection marker (*e.g., Neo^r*) and screened for non-random integration, can be further selected for multiple copies of the selectable marker gene by exposure to elevated levels of the selective agent (*e.g., G418*). The cells are then analyzed for homozygosity at the target locus. Alternatively, a second construct can be generated with a different positive selection marker inserted between the two homologous sequences. The two

constructs can be introduced into the cell either sequentially or simultaneously, followed by appropriate selection for each of the positive marker genes. The final cell is screened for homologous recombination of both alleles of the target.

Production of Transgenic Animals

5 Selected cells are then injected into a blastocyst (or other stage of development suitable for the purposes of creating a viable animal, such as, for example, a morula) of an animal (*e.g.*, a mouse) to form chimeras (*see e.g.*, Bradley, A. in *Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed., IRL, Oxford, pp. 113-152 (1987)). Alternatively, selected ES cells can be allowed to aggregate with
10 dissociated mouse embryo cells to form the aggregation chimera. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Chimeric progeny harbouring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA. In one embodiment, chimeric progeny mice are used to
15 generate a mouse with a heterozygous disruption in the CXCR6 gene. Heterozygous transgenic mice can then be mated. It is well known in the art that typically $\frac{1}{4}$ of the offspring of such matings will have a homozygous disruption in the CXCR6 gene.

 The heterozygous and homozygous transgenic mice can then be compared to normal, wild-type mice to determine whether disruption of the CXCR6 gene causes
20 phenotypic changes, especially pathological changes. For example, heterozygous and homozygous mice may be evaluated for phenotypic changes by physical examination, necropsy, histology, clinical chemistry, complete blood count, body weight, organ weights, and cytological evaluation of bone marrow. Phenotypic changes may also comprise behavioral modifications or abnormalities.

25 In one embodiment, the phenotype (or phenotypic change) associated with a disruption in the CXCR6 gene is placed into or stored in a database. Preferably, the database includes: (i) genotypic data (*e.g.*, identification of the disrupted gene) and (ii) phenotypic data (*e.g.*, phenotype(s) resulting from the gene disruption) associated with the genotypic data. The database is preferably electronic. In addition, the database is
30 preferably combined with a search tool so that the database is searchable.

Conditional Transgenic Animals

The present invention further contemplates conditional transgenic or knockout animals, such as those produced using recombination methods. Bacteriophage P1 Cre recombinase and flp recombinase from yeast plasmids are two non-limiting examples of site-specific DNA recombinase enzymes that cleave DNA at specific target sites (lox P sites for cre recombinase and frt sites for flp recombinase) and catalyze a ligation of this DNA to a second cleaved site. A large number of suitable alternative site-specific recombinases have been described, and their genes can be used in accordance with the method of the present invention. Such recombinases include the Int recombinase of bacteriophage λ (with or without Xis) (Weisberg, R. *et al.*, in *Lambda II*, (Hendrix, R., *et al.*, Eds.), Cold Spring Harbor Press, Cold Spring Harbor, NY, pp. 211-50 (1983), herein incorporated by reference); TpnI and the β -lactamase transposons (Mercier, *et al.*, *J. Bacteriol.*, 172:3745-57 (1990)); the Tn3 resolvase (Flanagan & Fennewald *J. Molec. Biol.*, 206:295-304 (1989); Stark, *et al.*, *Cell*, 58:779-90 (1989)); the yeast recombinases (Matsuzaki, *et al.*, *J. Bacteriol.*, 172:610-18 (1990)); the *B. subtilis* SpoIVC recombinase (Sato, *et al.*, *J. Bacteriol.* 172:1092-98 (1990)); the Flp recombinase (Schwartz & Sadowski, *J. Molec. Biol.*, 205:647-658 (1989); Parsons, *et al.*, *J. Biol. Chem.*, 265:4527-33 (1990); Golic & Lindquist, *Cell*, 59:499-509 (1989); Amin, *et al.*, *J. Molec. Biol.*, 214:55-72 (1990)); the Hin recombinase (Glasgow, *et al.*, *J. Biol. Chem.*, 264:10072-82 (1989)); immunoglobulin recombinases (Malynn, *et al.*, *Cell*, 54:453-460 (1988)); and the Cin recombinase (Haffter & Bickle, *EMBO J.*, 7:3991-3996 (1988); Hubner, *et al.*, *J. Molec. Biol.*, 205:493-500 (1989)), all herein incorporated by reference. Such systems are discussed by Echols (*J. Biol. Chem.* 265:14697-14700 (1990)); de Villartay (*Nature*, 335:170-74 (1988)); Craig, (*Ann. Rev. Genet.*, 22:77-105 (1988)); Poyart-Salmeron, *et al.*, (*EMBO J.* 8:2425-33 (1989)); Hunger-Bertling, *et al.*, (*Mol Cell. Biochem.*, 92:107-16 (1990)); and Cregg & Madden (*Mol. Gen. Genet.*, 219:320-23 (1989)), all herein incorporated by reference.

Cre has been purified to homogeneity, and its reaction with the loxP site has been extensively characterized (Abremski & Hess *J. Mol. Biol.* 259:1509-14 (1984), herein incorporated by reference). Cre protein has a molecular weight of 35,000 and can be obtained commercially from New England Nuclear/Du Pont. The cre gene (which

encodes the Cre protein) has been cloned and expressed (Abremski, *et al.*, *Cell* 32:1301-11 (1983), herein incorporated by reference). The Cre protein mediates recombination between two loxP sequences (Sternberg, *et al.*, *Cold Spring Harbor Symp. Quant. Biol.* 45:297-309 (1981)), which may be present on the same or different DNA molecule. Because the internal spacer sequence of the loxP site is asymmetrical, two loxP sites can exhibit directionality relative to one another (Hoess & Abremski *Proc. Natl. Acad. Sci. U.S.A.* 81:1026-29 (1984)). Thus, when two sites on the same DNA molecule are in a directly repeated orientation, Cre will excise the DNA between the sites (Abremski, *et al.*, *Cell* 32:1301-11 (1983)). However, if the sites are inverted with respect to each other, the DNA between them is not excised after recombination but is simply inverted. Thus, a circular DNA molecule having two loxP sites in direct orientation will recombine to produce two smaller circles, whereas circular molecules having two loxP sites in an inverted orientation simply invert the DNA sequences flanked by the loxP sites. In addition, recombinase action can result in reciprocal exchange of regions distal to the target site when targets are present on separate DNA molecules.

Recombinases have important application for characterizing gene function in knockout models. When the constructs described herein are used to disrupt CXCR6 genes, a fusion transcript can be produced when insertion of the positive selection marker occurs downstream (3') of the translation initiation site of the CXCR6 gene. The fusion transcript could result in some level of protein expression with unknown consequence. It has been suggested that insertion of a positive selection marker gene can affect the expression of nearby genes. These effects may make it difficult to determine gene function after a knockout event since one could not discern whether a given phenotype is associated with the inactivation of a gene, or the transcription of nearby genes. Both potential problems are solved by exploiting recombinase activity. When the positive selection marker is flanked by recombinase sites in the same orientation, the addition of the corresponding recombinase will result in the removal of the positive selection marker. In this way, effects caused by the positive selection marker or expression of fusion transcripts are avoided.

In one embodiment, purified recombinase enzyme is provided to the cell by direct microinjection. In another embodiment, recombinase is expressed from a co-transfected

construct or vector in which the recombinase gene is operably linked to a functional promoter. An additional aspect of this embodiment is the use of tissue-specific or inducible recombinase constructs that allow the choice of when and where recombination occurs. One method for practicing the inducible forms of recombinase-mediated recombination involves the use of vectors that use inducible or tissue-specific promoters or other gene regulatory elements to express the desired recombinase activity. The inducible expression elements are preferably operatively positioned to allow the inducible control or activation of expression of the desired recombinase activity. Examples of such inducible promoters or other gene regulatory elements include, but are not limited to, tetracycline, metallothionine, ecdysone, and other steroid-responsive promoters, rapamycin responsive promoters, and the like (No, *et al.*, *Proc. Natl. Acad. Sci. USA*, 93:3346-51 (1996); Furth, *et al.*, *Proc. Natl. Acad. Sci. USA*, 91:9302-6 (1994)). Additional control elements that can be used include promoters requiring specific transcription factors such as viral, promoters. Vectors incorporating such promoters would only express recombinase activity in cells that express the necessary transcription factors.

Models for Disease

The cell- and animal-based systems described herein can be utilized as models for diseases. Animals of any species, including, but not limited to, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, goats, and non-human primates, *e.g.*, baboons, monkeys, and chimpanzees may be used to generate disease animal models. In addition, cells from humans may be used. These systems may be used in a variety of applications. Such assays may be utilized as part of screening strategies designed to identify agents, such as compounds that are capable of ameliorating disease symptoms. Thus, the animal- and cell-based models may be used to identify drugs, pharmaceuticals, therapies and interventions that may be effective in treating disease.

Cell-based systems may be used to identify compounds that may act to ameliorate disease symptoms. For example, such cell systems may be exposed to a compound suspected of exhibiting an ability to ameliorate disease symptoms, at a sufficient concentration and for a time sufficient to elicit such an amelioration of disease symptoms in the exposed cells. After exposure, the cells are examined to determine whether one or

more of the disease cellular phenotypes has been altered to resemble a more normal or more wild-type, non-disease phenotype.

In addition, animal-based disease systems, such as those described herein, may be used to identify compounds capable of ameliorating disease symptoms. Such animal models may be used as test substrates for the identification of drugs, pharmaceuticals, therapies, and interventions that may be effective in treating a disease or other phenotypic characteristic of the animal. For example, animal models may be exposed to a compound or agent suspected of exhibiting an ability to ameliorate disease symptoms, at a sufficient concentration and for a time sufficient to elicit such an amelioration of disease symptoms in the exposed animals. The response of the animals to the exposure may be monitored by assessing the reversal of disorders associated with the disease. Exposure may involve treating mother animals during gestation of the model animals described herein, thereby exposing embryos or fetuses to the compound or agent that may prevent or ameliorate the disease or phenotype. Neonatal, juvenile, and adult animals can also be exposed.

More particularly, using the animal models of the invention, methods of identifying agents are provided, in which such agents can be identified on the basis of their ability to affect at least one phenotype associated with a disruption in a CXCR6 gene. In one embodiment, the present invention provides a method of identifying agents having an effect on CXCR6 expression or function. The method includes measuring a physiological response of the animal, for example, to the agent and comparing the physiological response of such animal to a control animal, wherein the physiological response of the animal comprising a disruption in a CXCR6 as compared to the control animal indicates the specificity of the agent. A "physiological response" is any biological or physical parameter of an animal that can be measured. Molecular assays (*e.g.*, gene transcription, protein production and degradation rates), physical parameters (*e.g.*, exercise physiology tests, measurement of various parameters of respiration, measurement of heart rate or blood pressure and measurement of bleeding time), behavioral testing, and cellular assays (*e.g.*, immunohistochemical assays of cell surface markers, or the ability of cells to aggregate or proliferate) can be used to assess a physiological response.

The transgenic animals and cells of the present invention may be utilized as models for diseases, disorders, or conditions associated with phenotypes relating to a disruption in a CXCR6 gene.

The present invention provides a unique animal model for testing and developing new treatments relating to the behavioral phenotypes. Analysis of the behavioral phenotype allows for the development of an animal model useful for testing, for instance, the efficacy of proposed genetic and pharmacological therapies for human genetic diseases, such as neurological, neuropsychological, or psychotic illnesses.

A statistical analysis of the various behaviors measured can be carried out using any conventional statistical program routinely used by those skilled in the art (such as, for example, "Analysis of Variance" or ANOVA). A "p" value of about 0.05 or less is generally considered to be statistically significant, although slightly higher p values may still be indicative of statistically significant differences. To statistically analyze abnormal behavior, a comparison is made between the behavior of a transgenic animal (or a group thereof) to the behavior of a wild-type mouse (or a group thereof), typically under certain prescribed conditions. "Abnormal behavior" as used herein refers to behavior exhibited by an animal having a disruption in the CXCR6 gene, *e.g.* transgenic animal, which differs from an animal without a disruption in the CXCR6 gene, *e.g.* wild-type mouse. Abnormal behavior consists of any number of standard behaviors that can be objectively measured (or observed) and compared. In the case of comparison, it is preferred that the change be statistically significant to confirm that there is indeed a meaningful behavioral difference between the knockout animal and the wild-type control animal. Examples of behaviors that may be measured or observed include, but are not limited to, ataxia, rapid limb movement, eye movement, breathing, motor activity, cognition, emotional behaviors, social behaviors, hyperactivity, hypersensitivity, anxiety, impaired learning, abnormal reward behavior, and abnormal social interaction, such as aggression.

A series of tests may be used to measure the behavioral phenotype of the animal models of the present invention, including neurological and neuropsychological tests to identify abnormal behavior. These tests may be used to measure abnormal behavior relating to, for example, learning and memory, eating, pain, aggression, sexual

reproduction, anxiety, depression, schizophrenia, and drug abuse. (see, e.g., Crawley & Paylor, *Hormones and Behavior* 31:197-211 (1997)).

The social interaction test involves exposing a mouse to other animals in a variety of settings. The social behaviors of the animals (e.g., touching, climbing, sniffing, and mating) are subsequently evaluated. Differences in behaviors can then be statistically analyzed and compared (see, e.g., S. E. File, et al., *Pharmacol. Bioch. Behav.* 22:941-944 (1985); R. R. Holson, *Phys. Behav.* 37:239-247 (1986)). Exemplary behavioral tests include the following.

The mouse startle response test typically involves exposing the animal to a sensory (typically auditory) stimulus and measuring the startle response of the animal (see, e.g., M. A. Geyer, et al., *Brain Res. Bull.* 25:485-498 (1990); Paylor and Crawley, *Psychopharmacology* 132:169-180 (1997)). A pre-pulse inhibition test can also be used, in which the percent inhibition (from a normal startle response) is measured by “cueing” the animal first with a brief low-intensity pre-pulse prior to the startle pulse.

The electric shock test generally involves exposure to an electrified surface and measurement of subsequent behaviors such as, for example, motor activity, learning, social behaviors. The behaviors are measured and statistically analyzed using standard statistical tests. (see, e.g., G. J. Kant, et al., *Pharm. Bioch. Behav.* 20:793-797 (1984); N. J. Leidenheimer, et al., *Pharmacol. Bioch. Behav.* 30:351-355 (1988)).

The tail-pinch or immobilization test involves applying pressure to the tail of the animal and/or restraining the animal's movements. Motor activity, social behavior, and cognitive behavior are examples of the areas that are measured. (see, e.g., M. Bertolucci D'Angic, et al., *Neurochem.* 55:1208-1214 (1990)).

The novelty test generally comprises exposure to a novel environment and/or novel objects. The animal's motor behavior in the novel environment and/or around the novel object are measured and statistically analyzed. (see, e.g., D. K. Reinstein, et al., *Pharm. Bioch. Behav.* 17:193-202 (1982); B. Poucet, *Behav. Neurosci.* 103:1009-10016 (1989); R. R. Holson, et al., *Phys. Behav.* 37:231-238 (1986)). This test may be used to detect visual processing deficiencies or defects.

The learned helplessness test involves exposure to stresses, for example, noxious stimuli, which cannot be affected by the animal's behavior. The animal's behavior can be

statistically analyzed using various standard statistical tests. (*see, e.g., A. Leshner, et al., Behav. Neural Biol. 26:497-501 (1979)*).

Alternatively, a tail suspension test may be used, in which the “immobile” time of the mouse is measured when suspended “upside-down” by its tail. This is a measure of whether the animal struggles, an indicator of depression. In humans, depression is believed to result from feelings of a lack of control over one’s life or situation. It is believed that a depressive state can be elicited in animals by repeatedly subjecting them to aversive situations over which they have no control. A condition of “learned helplessness” is eventually reached, in which the animal will stop trying to change its circumstances and simply accept its fate. Animals that stop struggling sooner are believed to be more prone to depression. Studies have shown that the administration of certain antidepressant drugs prior to testing increases the amount of time that animals struggle before giving up.

The Morris water-maze test comprises learning spatial orientations in water and subsequently measuring the animal's behaviors, such as, for example, by counting the number of incorrect choices. The behaviors measured are statistically analyzed using standard statistical tests. (*see, e.g., E. M. Spruijt, et al., Brain Res. 527:192-197 (1990)*).

Alternatively, a Y-shaped maze may be used (*see, e.g., McFarland, D.J., Pharmacology, Biochemistry and Behavior 32:723-726 (1989); Dellu, F., et al., Neurobiology of Learning and Memory 73:31-48 (2000)*). The Y-maze is generally believed to be a test of cognitive ability. The dimensions of each arm of the Y-maze can be, for example, approximately 40 cm x 8 cm x 20 cm, although other dimensions may be used. Each arm can also have, for example, sixteen equally spaced photobeams to automatically detect movement within the arms. At least two different tests can be performed using such a Y-maze. In a continuous Y-maze paradigm, mice are allowed to explore all three arms of a Y-maze for, *e.g., approximately 10 minutes*. The animals are continuously tracked using photobeam detection grids, and the data can be used to measure spontaneous alternation and positive bias behavior. Spontaneous alternation refers to the natural tendency of a “normal” animal to visit the least familiar arm of a maze. An alternation is scored when the animal makes two consecutive turns in the same direction, thus representing a sequence of visits to the least recently entered arm of the maze.

Position bias determines egocentrically defined responses by measuring the animal's tendency to favor turning in one direction over another. Therefore, the test can detect differences in an animal's ability to navigate on the basis of allocentric or egocentric mechanisms. The two-trial Y-maze memory test measures response to novelty and spatial memory based on a free-choice exploration paradigm. During the first trial (acquisition), the animals are allowed to freely visit two arms of the Y-maze for, *e.g.*, approximately 15 minutes. The third arm is blocked off during this trial. The second trial (retrieval) is performed after an intertrial interval of, *e.g.*, approximately 2 hours. During the retrieval trial, the blocked arm is opened and the animal is allowed access to all three arms for, *e.g.*, approximately 5 minutes. Data are collected during the retrieval trial and analyzed for the number and duration of visits to each arm. Because the three arms of the maze are virtually identical, discrimination between novelty and familiarity is dependent on "environmental" spatial cues around the room relative to the position of each arm. Changes in arm entry and duration of time spent in the novel arm in a transgenic animal model may be indicative of a role of that gene in mediating novelty and recognition processes.

The passive avoidance or shuttle box test generally involves exposure to two or more environments, one of which is noxious, providing a choice to be learned by the animal. Behavioral measures include, for example, response latency, number of correct responses, and consistency of response. (*see, e.g., R. Ader, et al., Psychon. Sci.* 26:125-128 (1972); R. R. Holson, *Phys. Behav.* 37:221-230 (1986)). Alternatively, a zero-maze can be used. In a zero-maze, the animals can, for example, be placed in a closed quadrant of an elevated annular platform having, *e.g.*, 2 open and 2 closed quadrants, and are allowed to explore for approximately 5 minutes. This paradigm exploits an approach-avoidance conflict between normal exploratory activity and an aversion to open spaces in rodents. This test measures anxiety levels and can be used to evaluate the effectiveness of anti-anxiolytic drugs. The time spent in open quadrants versus closed quadrants may be recorded automatically, with, for example, the placement of photobeams at each transition site.

The food avoidance test involves exposure to novel food and objectively measuring, for example, food intake and intake latency. The behaviors measured are

statistically analyzed using standard statistical tests. (see, e.g., B. A. Campbell, *et al.*, *J. Comp. Physiol. Psychol.* 67:15-22 (1969)).

The elevated plus-maze test comprises exposure to a maze, without sides, on a platform, the animal's behavior is objectively measured by counting the number of maze entries and maze learning. The behavior is statistically analyzed using standard statistical tests. (see, e.g., H. A. Baldwin, *et al.*, *Brain Res. Bull.* 20:603-606 (1988)).

The stimulant-induced hyperactivity test involves injection of stimulant drugs (e.g., amphetamines, cocaine, PCP, and the like), and objectively measuring, for example, motor activity, social interactions, cognitive behavior. The animal's behaviors are statistically analyzed using standard statistical tests. (see, e.g., P. B. S. Clarke, *et al.*, *Psychopharmacology* 96:511-520 (1988); P. Kuczenski, *et al.*, *J. Neuroscience* 11:2703-2712 (1991)).

The self-stimulation test generally comprises providing the mouse with the opportunity to regulate electrical and/or chemical stimuli to its own brain. Behavior is measured by frequency and pattern of self-stimulation. Such behaviors are statistically analyzed using standard statistical tests. (see, e.g., S. Nassif, *et al.*, *Brain Res.*, 332:247-257 (1985); W. L. Isaac, *et al.*, *Behav. Neurosci.* 103:345-355 (1989)).

The reward test involves shaping a variety of behaviors, e.g., motor, cognitive, and social, measuring, for example, rapidity and reliability of behavioral change, and statistically analyzing the behaviors measured. (see, e.g., L. E. Jarrard, *et al.*, *Exp. Brain Res.* 61:519-530 (1986)).

The DRL (differential reinforcement to low rates of responding) performance test involves exposure to intermittent reward paradigms and measuring the number of proper responses, e.g., lever pressing. Such behavior is statistically analyzed using standard statistical tests. (see, e.g., J. D. Sinden, *et al.*, *Behav. Neurosci.* 100:320-329 (1986); V. Nalwa, *et al.*, *Behav Brain Res.* 17:73-76 (1985); and A. J. Nonneman, *et al.*, *J. Comp. Physiol. Psych.* 95:588-602 (1981)).

The spatial learning test involves exposure to a complex novel environment, measuring the rapidity and extent of spatial learning, and statistically analyzing the behaviors measured. (see, e.g., N. Pitsikas, *et al.*, *Pharm. Bioch. Behav.* 38:931-934 (1991); B. Poucet, *et al.*, *Brain Res.* 37:269-280 (1990); D. Christie, *et al.*, *Brain Res.*

37:263-268 (1990); and F. Van Haaren, *et al.*, Behav. Neurosci. 102:481-488 (1988)).

Alternatively, an open-field (of) test may be used, in which the greater distance traveled for a given amount of time is a measure of the activity level and anxiety of the animal.

When the open field is a novel environment, it is believed that an approach-avoidance

5 situation is created, in which the animal is “torn” between the drive to explore and the drive to protect itself. Because the chamber is lighted and has no places to hide other than the corners, it is expected that a “normal” mouse will spend more time in the corners and around the periphery than it will in the center where there is no place to hide.

“Normal” mice will, however, venture into the central regions as they explore more and

10 more of the chamber. It can then be extrapolated that especially anxious mice will spend most of their time in the corners, with relatively little or no exploration of the central region, whereas bold (*i.e.*, less anxious) mice will travel a greater distance, showing little preference for the periphery versus the central region.

The visual, somatosensory and auditory neglect tests generally comprise exposure
15 to a sensory stimulus, objectively measuring, for example, orientating responses, and statistically analyzing the behaviors measured. (*see, e.g.*, J. M. Vargo, *et al.*, Exp. Neurol. 102:199-209 (1988)).

The consummatory behavior test generally comprises feeding and drinking, and objectively measuring quantity of consumption. The behavior measured is statistically
20 analyzed using standard statistical tests. (*see, e.g.*, P. J. Fletcher, *et al.*, Psychopharmacol. 102:301-308 (1990); M. G. Corda, *et al.*, Proc. Nat'l Acad. Sci. USA 80:2072-2076 (1983)).

A visual discrimination test can also be used to evaluate the visual processing of an animal. One or two similar objects are placed in an open field and the animal is
25 allowed to explore for about 5-10 minutes. The time spent exploring each object (proximity to, *i.e.*, movement within, *e.g.*, about 3-5 cm of the object is considered exploration of an object) is recorded. The animal is then removed from the open field, and the objects are replaced by a similar object and a novel object. The animal is returned to the open field and the percent time spent exploring the novel object over the
30 old object is measured (again, over about a 5-10 minute span). “Normal” animals will typically spend a higher percentage of time exploring the novel object rather than the old

object. If a delay is imposed between sampling and testing, the memory task becomes more hippocampal-dependent. If no delay is imposed, the task is more based on simple visual discrimination. This test can also be used for olfactory discrimination, in which the objects (preferably, simple blocks) can be sprayed or otherwise treated to hold an odor. This test can also be used to determine if the animal can make gustatory discriminations; animals that return to the previously eaten food instead of novel food exhibit gustatory neophobia.

A hot plate analgesia test can be used to evaluate an animal's sensitivity to heat or painful stimuli. For example, a mouse can be placed on an approximately 55°C hot plate and the mouse's response latency (*e.g.*, time to pick up and lick a hind paw) can be recorded. These responses are not reflexes, but rather "higher" responses requiring cortical involvement. This test may be used to evaluate a nociceptive disorder.

A tail-flick test may also be used to evaluate an animal's sensitivity to heat or painful stimuli. For example, a high-intensity thermal stimulus can be directed to the tail of a mouse and the mouse's response latency recorded (*e.g.*, the time from onset of stimulation to a rapid flick/withdrawal from the heat source) can be recorded. These responses are simple nociceptive reflexive responses that are involuntary spinally mediated flexion reflexes. This test may also be used to evaluate a nociceptive disorder.

An accelerating rotarod test may be used to measure coordination and balance in mice. Animals can be, for example, placed on a rod that acts like a rotating treadmill (or rolling log). The rotarod can be made to rotate slowly at first and then progressively faster until it reaches a speed of, *e.g.*, approximately 60 rpm. The mice must continually reposition themselves in order to avoid falling off. The animals are preferably tested in at least three trials, a minimum of 20 minutes apart. Those mice that are able to stay on the rod the longest are believed to have better coordination and balance.

A metrazol administration test can be used to screen animals for varying susceptibilities to seizures or similar events. For example, a 5mg/ml solution of metrazol can be infused through the tail vein of a mouse at a rate of, *e.g.*, approximately 0.375 ml/min. The infusion will cause all mice to experience seizures, followed by death. Those mice that enter the seizure stage the soonest are believed to be more prone to seizures. Four distinct physiological stages can be recorded: soon after the start of

infusion, the mice will exhibit a noticeable “twitch”, followed by a series of seizures, ending in a final tensing of the body known as “tonic extension”, which is followed by death.

CXCR6 Gene Products

5 The present invention further contemplates use of the CXCR6 gene sequence to produce CXCR6 gene products. CXCR6 gene products may include proteins that represent functionally equivalent gene products. Such an equivalent gene product may contain deletions, additions or substitutions of amino acid residues within the amino acid sequence encoded by the gene sequences described herein, but which result in a silent
10 change, thus producing a functionally equivalent CXCR6 gene product. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. CXCR6 nucleic acid sequences and amino acid sequences may include the sequence shown in Figure 1 (SEQ ID NO:1) or identified in GenBank as Accession No.:
15 AF301018; GI:11342661; the CXCR6 polypeptide as shown in Figure 2 (SEQ ID NO:2) or identified in GenBank Accession No.: AAG34367; GI:11342662; or any homologues, orthologs, variants, derivatives, active fragments or mutants of CXCR6.

For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral
20 amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid. "Functionally equivalent", as utilized herein, refers to a protein capable of exhibiting a substantially similar *in vivo* activity as the endogenous gene products encoded by the
25 CXCR6 gene sequences. Alternatively, when utilized as part of an assay, "functionally equivalent" may refer to peptides capable of interacting with other cellular or extracellular molecules in a manner substantially similar to the way in which the corresponding portion of the endogenous gene product would.

Other protein products useful according to the methods of the invention are
30 peptides derived from or based on the CXCR6 gene products produced by recombinant or synthetic means (derived peptides).

CXCR6 gene products may be produced by recombinant DNA technology using techniques well known in the art. Thus, methods for preparing the CXCR6 polypeptides and peptides of the invention by expressing nucleic acids encoding gene sequences are described herein. Methods that are well known to those skilled in the art can be used to
5 construct expression vectors containing CXCR6 protein coding sequences and appropriate transcriptional/translational control signals. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic techniques and *in vivo* recombination/genetic recombination (*see, e.g.*, Sambrook, *et al.*, 1989, *supra*, and Ausubel, *et al.*, 1989, *supra*). Alternatively, RNA capable of encoding CXCR6 protein
10 sequences may be chemically synthesized using, for example, automated synthesizers (*see, e.g.* Oligonucleotide Synthesis: A Practical Approach, Gait, M. J. ed., IRL Press, Oxford (1984)).

A variety of host-expression vector systems may be utilized to express the CXCR6 coding sequences of the invention. Such host-expression systems represent
15 vehicles by which the coding sequences of interest may be produced and subsequently purified, but also represent cells that may, when transformed or transfected with the appropriate nucleotide coding sequences, exhibit the CXCR6 protein of the invention *in situ*. These include but are not limited to microorganisms such as bacteria (*e.g.*, *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid
20 DNA expression vectors containing CXCR6 protein coding sequences; yeast (*e.g.* *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing the CXCR6 protein coding sequences; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing the CXCR6 protein coding sequences; plant cell systems infected with recombinant virus expression vectors
25 (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing CXCR6 protein coding sequences; or mammalian cell systems (*e.g.* COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (*e.g.*, metallothioneine promoter) or from mammalian viruses
30 (*e.g.*, the adenovirus late promoter; the vaccinia virus 7.5 K promoter).

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the CXCR6 protein being expressed. For example, when a large quantity of such a protein is to be produced, for the generation of antibodies or to screen peptide libraries, for example, vectors that direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not limited, to the *E. coli* expression vector pUR278 (Ruther *et al.*, *EMBO J.*, 2:1791-94 (1983)), in which the CXCR6 protein coding sequence may be ligated individually into the vector in frame with the *lac Z* coding region so that a fusion protein is produced; pIN vectors (Inouye & Inouye, *Nucleic Acids Res.*, 13:3101-09 (1985); Van Heeke *et al.*, *J. Biol. Chem.*, 264:5503-9 (1989)); and the like. pGEX vectors may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned CXCR6 protein can be released from the GST moiety.

In a preferred embodiment, full length cDNA sequences are appended with in-frame Bam HI sites at the amino terminus and Eco RI sites at the carboxyl terminus using standard PCR methodologies (Innis, *et al.* (eds) PCR Protocols: A Guide to Methods and Applications, Academic Press, San Diego (1990)) and ligated into the pGEX-2TK vector (Pharmacia, Uppsala, Sweden). The resulting cDNA construct contains a kinase recognition site at the amino terminus for radioactive labeling and glutathione S-transferase sequences at the carboxyl terminus for affinity purification (Nilsson, *et al.*, *EMBO J.*, 4: 1075-80 (1985); Zabeau *et al.*, *EMBO J.*, 1: 1217-24 (1982)).

In an insect system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes. The virus grows in *Spodoptera frugiperda* cells. The gene coding sequence may be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter). Successful insertion of gene coding sequence will result in inactivation of the polyhedrin gene and production of non-occluded recombinant virus (*i.e.*, virus lacking the proteinaceous coat coded for by the

polyhedrin gene). These recombinant viruses are then used to infect *Spodoptera frugiperda* cells in which the inserted gene is expressed (see, e.g., Smith, *et al.*, *J. Virol.* 46: 584-93 (1983); U.S. Patent No. 4,745,051).

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the gene coding sequence of interest may be ligated to an adenovirus transcription/translation control complex, e.g., the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by *in vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (e.g., region E1 or E3) will result in a recombinant virus that is viable and capable of expressing CXCR6 protein in infected hosts. (e.g., see Logan *et al.*, *Proc. Natl. Acad. Sci. USA*, 81:3655-59 (1984)). Specific initiation signals may also be required for efficient translation of inserted gene coding sequences. These signals include the ATG initiation codon and adjacent sequences. In cases where an entire gene, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of the gene coding sequence is inserted, exogenous translational control signals, including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (see Bitter, *et al.*, *Methods in Enzymol.*, 153:516-44 (1987)).

In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells that possess the cellular machinery for proper processing of the

primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, etc.

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines that stably express the CXCR6 protein may be engineered. Rather than using expression vectors that contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (e.g., promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells that stably integrate the plasmid into their chromosomes and grow, to form foci, which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines that express the CXCR6 protein. Such engineered cell lines may be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the CXCR6 protein.

In a preferred embodiment, timing and/or quantity of expression of the recombinant protein can be controlled using an inducible expression construct. Inducible constructs and systems for inducible expression of recombinant proteins will be well known to those skilled in the art. Examples of such inducible promoters or other gene regulatory elements include, but are not limited to, tetracycline, metallothionine, ecdysone, and other steroid-responsive promoters, rapamycin responsive promoters, and the like (No, *et al.*, *Proc. Natl. Acad. Sci. USA*, 93:3346-51 (1996); Furth, *et al.*, *Proc. Natl. Acad. Sci. USA*, 91:9302-6 (1994)). Additional control elements that can be used include promoters requiring specific transcription factors such as viral, particularly HIV, promoters. In one embodiment, a Tet inducible gene expression system is utilized. (Gossen *et al.*, *Proc. Natl. Acad. Sci. USA*, 89:5547-51 (1992); Gossen, *et al.*, *Science*, 268:1766-69 (1995)). Tet Expression Systems are based on two regulatory elements derived from the tetracycline-resistance operon of the *E. coli* Tn10 transposon—the tetracycline repressor protein (TetR) and the tetracycline operator sequence (*tetO*) to which TetR binds. Using such a system, expression of the recombinant protein is placed

under the control of the *tetO* operator sequence and transfected or transformed into a host cell. In the presence of TetR, which is co-transfected into the host cell, expression of the recombinant protein is repressed due to binding of the TetR protein to the *tetO* regulatory element. High-level, regulated gene expression can then be induced in response to
5 varying concentrations of tetracycline (Tc) or Tc derivatives such as doxycycline (Dox), which compete with *tetO* elements for binding to TetR. Constructs and materials for tet inducible gene expression are available commercially from CLONTECH Laboratories, Inc., Palo Alto, CA.

When used as a component in an assay system, the CXCR6 protein may be
10 labeled, either directly or indirectly, to facilitate detection of a complex formed between the CXCR6 protein and a test substance. Any of a variety of suitable labeling systems may be used including but not limited to radioisotopes such as ¹²⁵I; enzyme labeling systems that generate a detectable calorimetric signal or light when exposed to substrate; and fluorescent labels. Where recombinant DNA technology is used to produce the
15 CXCR6 protein for such assay systems, it may be advantageous to engineer fusion proteins that can facilitate labeling, immobilization and/or detection.

Indirect labeling involves the use of a protein, such as a labeled antibody, which specifically binds to the gene product. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments and fragments produced
20 by a Fab expression library.

Production of Antibodies

Described herein are methods for the production of antibodies capable of specifically recognizing one or more epitopes. Such antibodies may include, but are not limited to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric
25 antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above. Such antibodies may be used, for example, in the detection of a CXCR6 gene in a biological sample, or, alternatively, as a method for the inhibition of abnormal CXCR6 gene activity. Thus, such antibodies may be utilized as
30 part of disease treatment methods, and/or may be used as part of diagnostic techniques

whereby patients may be tested for abnormal levels of CXCR6 proteins, or for the presence of abnormal forms of such proteins.

For the production of antibodies, various host animals may be immunized by injection with the CXCR6 gene, its expression product or a portion thereof. Such host animals may include but are not limited to rabbits, mice, rats, goats and chickens, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG (*bacille Calmette-Guerin*) and *Corynebacterium parvum*.

Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of animals immunized with an antigen, such as CXCR6 gene product, or an antigenic functional derivative thereof. For the production of polyclonal antibodies, host animals such as those described above, may be immunized by injection with gene product supplemented with adjuvants as also described above.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique that provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to the hybridoma technique of Köhler and Milstein, *Nature*, 256:495-7 (1975); and U.S. Patent No. 4,376,110), the human B-cell hybridoma technique (Kosbor, *et al.*, *Immunology Today*, 4:72 (1983); Cote, *et al.*, *Proc. Natl. Acad. Sci. USA*, 80:2026-30 (1983)), and the EBV-hybridoma technique (Cole, *et al.*, in *Monoclonal Antibodies And Cancer Therapy*, Alan R. Liss, Inc., New York, pp. 77-96 (1985)). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison, *et al.*, *Proc. Natl. Acad. Sci.*, 81:6851-6855 (1984); Takeda, *et al.*, *Nature*, 314:452-54 (1985)) by splicing the genes from a mouse antibody molecule of appropriate

antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region.

5 Alternatively, techniques described for the production of single chain antibodies (U.S. Patent No. 4,946,778; Bird, *Science* 242:423-26 (1988); Huston, *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:5879-83 (1988); and Ward, *et al.*, *Nature*, 334:544-46 (1989)) can be adapted to produce gene-single chain antibodies. Single chain antibodies are typically formed by linking the heavy and light chain fragments of the Fv region via an amino acid
10 bridge, resulting in a single chain polypeptide.

Antibody fragments that recognize specific epitopes may be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')₂ fragments that can be produced by pepsin digestion of the antibody molecule and the Fab fragments that can be generated by reducing the disulfide bridges of the F(ab')₂
15 fragments. Alternatively, Fab expression libraries may be constructed (Huse, *et al.*, *Science*, 246:1275-81 (1989)) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Screening Methods

Various animal-derived "preparations," including cells and tissues, as well as cell-
20 free extracts, homogenates, fractions and purified proteins, may be used to determine whether a particular agent is capable of modulating an activity of a CXCR6 gene product or a phenotype associated therewith. For example, such preparations may be generated according to methods well known in the art from the tissues or organs of wild-type and knockout animals. Wild-type, but not knockout, preparations will contain endogenous
25 CXCR6 gene product, as well as the native activities, interactions and effects of the CXCR6 gene product. Thus, when knockout and wild-type preparations are contacted with a test agent in parallel, the ability of the test agent to modulate CXCR6 gene product, or a phenotype associated therewith, can be determined. Agents capable of modulating an activity of a CXCR6 gene product or a phenotype associated therewith are
30 identified as those that modulate wild-type, but not knockout, preparations. Modulation may be detected, for example, as the ability of the agent to interact with a preparation,

thereby indicating interaction with the gene product itself or a product thereof.

Alternatively, the agent may affect a structural, metabolic or biochemical feature of the preparation, such as enzymatic activity of the preparation related to the CXCR6 gene product. An inclusive discussion of the events for which modulation by a test agent may be observed is beyond the scope of this application, but will be well known by those skilled in the art.

The present invention may be employed in a process for screening for agents such as agonists, *i.e.*, agents that bind to and activate CXCR6 polypeptides, or antagonists, *i.e.*, inhibit the activity or interaction of CXCR6 polypeptides with its ligand. Thus, polypeptides of the invention may also be used to assess the binding of small molecule substrates and ligands in, for example, cells, cell-free preparations, chemical libraries, and natural product mixtures as known in the art. Any methods routinely used to identify and screen for agents that can modulate receptors may be used in accordance with the present invention.

The present invention provides methods for identifying and screening for agents that modulate CXCR6 expression or function. More particularly, cells that contain and express CXCR6 gene sequences may be used to screen for therapeutic agents. Such cells may include non-recombinant monocyte cell lines, such as U937 (ATCC# CRL-1593), THP-1 (ATCC# TIB-202), and P388D1 (ATCC# TIB-63); endothelial cells such as HUVEC's and bovine aortic endothelial cells (BAEC's); as well as generic mammalian cell lines such as HeLa cells and COS cells, *e.g.*, COS-7 (ATCC# CRL-1651). Further, such cells may include recombinant, transgenic cell lines. For example, the transgenic mice of the invention may be used to generate cell lines, containing one or more cell types involved in a disease, that can be used as cell culture models for that disorder.

While cells, tissues, and primary cultures derived from the disease transgenic animals of the invention may be utilized, the generation of continuous cell lines is preferred. For examples of techniques that may be used to derive a continuous cell line from the transgenic animals, see Small, *et al.*, *Mol. Cell Biol.*, 5:642-48 (1985).

CXCR6 gene sequences may be introduced into and overexpressed in, the genome of the cell of interest. In order to overexpress a CXCR6 gene sequence, the coding portion of the CXCR6 gene sequence may be ligated to a regulatory sequence that is

capable of driving gene expression in the cell type of interest. Such regulatory regions will be well known to those of skill in the art, and may be utilized in the absence of undue experimentation. CXCR6 gene sequences may also be disrupted or underexpressed.

Cells having CXCR6 gene disruptions or underexpressed CXCR6 gene sequences may be used, for example, to screen for agents capable of affecting alternative pathways that compensate for any loss of function attributable to the disruption or underexpression.

In vitro systems may be designed to identify compounds capable of binding the CXCR6 gene products. Such compounds may include, but are not limited to, peptides made of D-and/or L-configuration amino acids (in, for example, the form of random peptide libraries; (see e.g., Lam, *et al.*, *Nature*, 354:82-4 (1991)), phosphopeptides (in, for example, the form of random or partially degenerate, directed phosphopeptide libraries; see, e.g., Songyang, *et al.*, *Cell*, 72:767-78 (1993)), antibodies, and small organic or inorganic molecules. Compounds identified may be useful, for example, in modulating the activity of CXCR6 proteins, preferably mutant CXCR6 proteins; elaborating the biological function of the CXCR6 protein; or screening for compounds that disrupt normal CXCR6 gene interactions or themselves disrupt such interactions.

The principle of the assays used to identify compounds that bind to the CXCR6 protein involves preparing a reaction mixture of the CXCR6 protein and the test compound under conditions and for a time sufficient to allow the two components to interact and bind, thus forming a complex that can be removed and/or detected in the reaction mixture. These assays can be conducted in a variety of ways. For example, one method to conduct such an assay would involve anchoring the CXCR6 protein or the test substance onto a solid phase and detecting target protein/test substance complexes anchored on the solid phase at the end of the reaction. In one embodiment of such a method, the CXCR6 protein may be anchored onto a solid surface, and the test compound, which is not anchored, may be labeled, either directly or indirectly.

In practice, microtitre plates are conveniently utilized. The anchored component may be immobilized by non-covalent or covalent attachments. Non-covalent attachment may be accomplished simply by coating the solid surface with a solution of the protein and drying. Alternatively, an immobilized antibody, preferably a monoclonal antibody,

specific for the protein may be used to anchor the protein to the solid surface. The surfaces may be prepared in advance and stored.

In order to conduct the assay, the nonimmobilized component is added to the coated surface containing the anchored component. After the reaction is complete, unreacted components are removed (*e.g.*, by washing) under conditions such that any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways. Where the previously nonimmobilized component is pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the previously nonimmobilized component is not pre-labeled, an indirect label can be used to detect complexes anchored on the surface; *e.g.*, using a labeled antibody specific for the previously nonimmobilized component (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody).

Alternatively, a reaction can be conducted in a liquid phase, the reaction products separated from unreacted components, and complexes detected; *e.g.*, using an immobilized antibody specific for CXCR6 gene product or the test compound to anchor any complexes formed in solution, and a labeled antibody specific for the other component of the possible complex to detect anchored complexes.

Compounds that are shown to bind to a particular CXCR6 gene product through one of the methods described above can be further tested for their ability to elicit a biochemical response from the CXCR6 protein. Agonists, antagonists and/or inhibitors of the expression product can be identified utilizing assays well known in the art.

Antisense, Ribozymes, and Antibodies

Other agents that may be used as therapeutics include the CXCR6 gene, its expression product(s) and functional fragments thereof. Additionally, agents that reduce or inhibit mutant CXCR6 gene activity may be used to ameliorate disease symptoms. Such agents include antisense, ribozyme, and triple helix molecules. Techniques for the production and use of such molecules are well known to those of skill in the art.

Anti-sense RNA and DNA molecules act to directly block the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. With respect to antisense DNA, oligodeoxyribonucleotides derived from the translation

initiation site, *e.g.*, between the -10 and +10 regions of the CXCR6 gene nucleotide sequence of interest, are preferred.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by an endonucleolytic cleavage. The composition of ribozyme molecules must include one or more sequences complementary to the CXCR6 gene mRNA, and must include the well known catalytic sequence responsible for mRNA cleavage. For this sequence, see U.S. Patent No. 5,093,246, which is incorporated by reference herein in its entirety. As such within the scope of the invention are engineered hammerhead motif ribozyme molecules that specifically and efficiently catalyze endonucleolytic cleavage of RNA sequences encoding CXCR6 proteins.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the molecule of interest for ribozyme cleavage sites that include the following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the CXCR6 gene containing the cleavage site may be evaluated for predicted structural features, such as secondary structure, that may render the oligonucleotide sequence unsuitable. The suitability of candidate sequences may also be evaluated by testing their accessibility to hybridization with complementary oligonucleotides, using ribonuclease protection assays.

Nucleic acid molecules to be used in triple helix formation for the inhibition of transcription should be single stranded and composed of deoxyribonucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Nucleotide sequences may be pyrimidine-based, which will result in TAT and CGC triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich molecules provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition, nucleic acid molecules may be chosen that are purine-rich, for example, containing a stretch of G residues. These molecules will form a

triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex.

Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so called "switchback" nucleic acid molecule. Switchback molecules are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

It is possible that the antisense, ribozyme, and/or triple helix molecules described herein may reduce or inhibit the transcription (triple helix) and/or translation (antisense, ribozyme) of mRNA produced by both normal and mutant CXCR6 gene alleles. In order to ensure that substantially normal levels of CXCR6 gene activity are maintained, nucleic acid molecules that encode and express CXCR6 polypeptides exhibiting normal activity may be introduced into cells that do not contain sequences susceptible to whatever antisense, ribozyme, or triple helix treatments are being utilized. Alternatively, it may be preferable to coadminister normal CXCR6 protein into the cell or tissue in order to maintain the requisite level of cellular or tissue CXCR6 gene activity.

Anti-sense RNA and DNA, ribozyme, and triple helix molecules of the invention may be prepared by any method known in the art for the synthesis of DNA and RNA molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides and oligoribonucleotides well known in the art such as for example solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors that incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into cell lines.

Various well-known modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but are not limited to the addition of flanking sequences of ribonucleotides or

deoxyribonucleotides to the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

Antibodies that are both specific for CXCR6 protein, and in particular, the mutant CXCR6 protein, and interfere with its activity may be used to inhibit mutant CXCR6 gene function. Such antibodies may be generated against the proteins themselves or against peptides corresponding to portions of the proteins using standard techniques known in the art and as also described herein. Such antibodies include but are not limited to polyclonal, monoclonal, Fab fragments, single chain antibodies, chimeric antibodies, antibody mimetics, etc.

In instances where the CXCR6 protein is intracellular and whole antibodies are used, internalizing antibodies may be preferred. However, lipofectin liposomes may be used to deliver the antibody or a fragment of the Fab region that binds to the CXCR6 gene epitope into cells. Where fragments of the antibody are used, the smallest inhibitory fragment that binds to the target or expanded target protein's binding domain is preferred. For example, peptides having an amino acid sequence corresponding to the domain of the variable region of the antibody that binds to the CXCR6 protein may be used. Such peptides may be synthesized chemically or produced via recombinant DNA technology using methods well known in the art (*see, e.g.,* Creighton, *Proteins: Structures and Molecular Principles* (1984) W.H. Freeman, New York 1983, *supra*; and Sambrook, *et al.*, 1989, *supra*). Alternatively, single chain neutralizing antibodies that bind to intracellular CXCR6 gene epitopes may also be administered. Such single chain antibodies may be administered, for example, by expressing nucleotide sequences encoding single-chain antibodies within the target cell population by utilizing, for example, techniques such as those described in Marasco, *et al.*, *Proc. Natl. Acad. Sci. USA*, 90:7889-93 (1993).

RNA sequences encoding CXCR6 protein may be directly administered to a patient exhibiting disease symptoms, at a concentration sufficient to produce a level of CXCR6 protein such that disease symptoms are ameliorated. Patients may be treated by gene replacement therapy. One or more copies of a normal CXCR6 gene, or a portion of the gene that directs the production of a normal CXCR6 protein with CXCR6 gene

function, may be inserted into cells using vectors that include, but are not limited to adenovirus, adeno-associated virus, and retrovirus vectors, in addition to other particles that introduce DNA into cells, such as liposomes. Additionally, techniques such as those described above may be utilized for the introduction of normal CXCR6 gene sequences into human cells.

Cells, preferably autologous cells, containing normal CXCR6 gene expressing gene sequences may then be introduced or reintroduced into the patient at positions that allow for the amelioration of disease symptoms.

Pharmaceutical Compositions, Effective Dosages, and Routes of Administration

The identified compounds that inhibit target mutant gene expression, synthesis and/or activity can be administered to a patient at therapeutically effective doses to treat or ameliorate the disease. A therapeutically effective dose refers to that amount of the compound sufficient to result in amelioration of symptoms of the disease.

Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, for determining the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD₅₀/ED₅₀. Compounds that exhibit large therapeutic indices are preferred. While compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC₅₀ (*i.e.*, the concentration of the test compound that achieves a half-maximal inhibition of symptoms) as determined in cell

culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

Pharmaceutical compositions for use in accordance with the present invention may be formulated in conventional manner using one or more physiologically acceptable carriers or excipients. Thus, the compounds and their physiologically acceptable salts and solvates may be formulated for administration by inhalation or insufflation (either through the mouth or the nose) or oral, buccal, parenteral, topical, subcutaneous, intraperitoneal, intravenous, intrapleural, intraocular, intraarterial, or rectal administration. It is also contemplated that pharmaceutical compositions may be administered with other products that potentiate the activity of the compound and optionally, may include other therapeutic ingredients.

For oral administration, the pharmaceutical compositions may take the form of, for example, tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (*e.g.*, pregelatinised maize starch, polyvinylpyrrolidone or hydroxypropyl methylcellulose); fillers (*e.g.*, lactose, microcrystalline cellulose or calcium hydrogen phosphate); lubricants (*e.g.*, magnesium stearate, talc or silica); disintegrants (*e.g.*, potato starch or sodium starch glycolate); or wetting agents (*e.g.*, sodium lauryl sulphate). The tablets may be coated by methods well known in the art. Liquid preparations for oral administration may take the form of, for example, solutions, syrups or suspensions, or they may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (*e.g.*, sorbitol syrup, cellulose derivatives or hydrogenated edible fats); emulsifying agents (*e.g.*, lecithin or acacia); non-aqueous vehicles (*e.g.*, almond oil, oily esters, ethyl alcohol or fractionated vegetable oils); and preservatives (*e.g.*, methyl or propyl-p-hydroxybenzoates or sorbic acid). The preparations may also contain buffer salts, flavoring, coloring and sweetening agents as appropriate.

Preparations for oral administration may be suitably formulated to give controlled release of the active compound.

For buccal administration the compositions may take the form of tablets or lozenges formulated in conventional manner.

For administration by inhalation, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant, *e.g.*, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of *e.g.* gelatin for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

The compounds may be formulated for parenteral administration by injection, *e.g.*, by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, *e.g.*, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, *e.g.*, sterile pyrogen-free water, before use.

The compounds may also be formulated in rectal compositions such as suppositories or retention enemas, *e.g.*, containing conventional suppository bases such as cocoa butter or other glycerides. Oral ingestion is possibly the easiest method of taking any medication. Such a route of administration, is generally simple and straightforward and is frequently the least inconvenient or unpleasant route of administration from the patient's point of view. However, this involves passing the material through the stomach, which is a hostile environment for many materials, including proteins and other biologically active compositions. As the acidic, hydrolytic and proteolytic environment of the stomach has evolved efficiently to digest proteinaceous materials into amino acids and oligopeptides for subsequent anabolism, it is hardly surprising that very little or any of a wide variety of biologically active proteinaceous material, if simply taken orally, would survive its passage through the stomach to be taken up by the body in the small intestine. The result, is that many proteinaceous medicaments must be taken in through

another method, such as parenterally, often by subcutaneous, intramuscular or intravenous injection.

Pharmaceutical compositions may also include various buffers (*e.g.*, Tris, acetate, phosphate), solubilizers (*e.g.*, Tween, Polysorbate), carriers such as human serum albumin, preservatives (thimerosal, benzyl alcohol) and anti-oxidants such as ascorbic acid in order to stabilize pharmaceutical activity. The stabilizing agent may be a detergent, such as tween-20, tween-80, NP-40 or Triton X-100. EBP may also be incorporated into particulate preparations of polymeric compounds for controlled delivery to a patient over an extended period of time. A more extensive survey of components in pharmaceutical compositions is found in Remington's Pharmaceutical Sciences, 18th ed., A. R. Gennaro, ed., Mack Publishing, Easton, Pa. (1990).

In addition to the formulations described previously, the compounds may also be formulated as a depot preparation. Such long acting formulations may be administered by implantation (for example, subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

The compositions may, if desired, be presented in a pack or dispenser device that may contain one or more unit dosage forms containing the active ingredient. The pack may for example comprise metal or plastic foil, such as a blister pack. The pack or dispenser device may be accompanied by instructions for administration.

Diagnostics

A variety of methods may be employed to diagnose disease conditions associated with the CXCR6 gene. Specifically, reagents may be used, for example, for the detection of the presence of CXCR6 gene mutations, or the detection of either over- or under-expression of CXCR6 gene mRNA.

According to the diagnostic and prognostic method of the present invention, alteration of the wild-type CXCR6 gene locus is detected. In addition, the method can be performed by detecting the wild-type CXCR6 gene locus and confirming the lack of a predisposition or neoplasia. "Alteration of a wild-type gene" encompasses all forms of

mutations including deletions, insertions and point mutations in the coding and noncoding regions. Deletions may be of the entire gene or only a portion of the gene. Point mutations may result in stop codons, frameshift mutations or amino acid substitutions. Somatic mutations are those that occur only in certain tissues, *e.g.*, in tumor tissue, and are not inherited in the germline. Germline mutations can be found in any of a body's tissues and are inherited. If only a single allele is somatically mutated, an early neoplastic state may be indicated. However, if both alleles are mutated, then a late neoplastic state may be indicated. The finding of gene mutations thus provides both diagnostic and prognostic information. A CXCR6 gene allele that is not deleted (*e.g.*, that found on the sister chromosome to a chromosome carrying a CXCR6 gene deletion) can be screened for other mutations, such as insertions, small deletions, and point mutations. Mutations found in tumor tissues may be linked to decreased expression of the CXCR6 gene product. However, mutations leading to non-functional gene products may also be linked to a cancerous state. Point mutational events may occur in regulatory regions, such as in the promoter of the gene, leading to loss or diminution of expression of the mRNA. Point mutations may also abolish proper RNA processing, leading to loss of expression of the CXCR6 gene product, or a decrease in mRNA stability or translation efficiency.

One test available for detecting mutations in a candidate locus is to directly compare genomic target sequences from cancer patients with those from a control population. Alternatively, one could sequence messenger RNA after amplification, *e.g.*, by PCR, thereby eliminating the necessity of determining the exon structure of the candidate gene. Mutations from cancer patients falling outside the coding region of the CXCR6 gene can be detected by examining the non-coding regions, such as introns and regulatory sequences near or within the CXCR6 gene. An early indication that mutations in noncoding regions are important may come from Northern blot experiments that reveal messenger RNA molecules of abnormal size or abundance in cancer patients as compared to control individuals.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one specific gene nucleic acid or anti-gene antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical

settings, to diagnose patients exhibiting disease symptoms or at risk for developing disease.

Any cell type or tissue, including brain, cortex, subcortical region, cerebellum, brainstem, olfactory bulb, spinal cord, eye, Harderian gland, heart, lung, liver, pancreas, kidney, spleen, thymus, lymph nodes, bone marrow, skin, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, large intestine, cecum, testis, epididymis, seminal vesicle, coagulating gland, prostate gland, ovary, uterus and white fat, in which the gene is expressed may be utilized in the diagnostics described below.

DNA or RNA from the cell type or tissue to be analyzed may easily be isolated using procedures that are well known to those in the art. Diagnostic procedures may also be performed *in situ* directly upon tissue sections (fixed and/or frozen) of patient tissue obtained from biopsies or resections, such that no nucleic acid purification is necessary. Nucleic acid reagents may be used as probes and/or primers for such *in situ* procedures (see, for example, Nuovo, PCR *In Situ* Hybridization: Protocols and Applications, Raven Press, N.Y. (1992)).

Gene nucleotide sequences, either RNA or DNA, may, for example, be used in hybridization or amplification assays of biological samples to detect disease-related gene structures and expression. Such assays may include, but are not limited to, Southern or Northern analyses, restriction fragment length polymorphism assays, single stranded conformational polymorphism analyses, *in situ* hybridization assays, and polymerase chain reaction analyses. Such analyses may reveal both quantitative aspects of the expression pattern of the gene, and qualitative aspects of the gene expression and/or gene composition. That is, such aspects may include, for example, point mutations, insertions, deletions, chromosomal rearrangements, and/or activation or inactivation of gene expression.

Preferred diagnostic methods for the detection of gene-specific nucleic acid molecules may involve for example, contacting and incubating nucleic acids, derived from the cell type or tissue being analyzed, with one or more labeled nucleic acid reagents under conditions favorable for the specific annealing of these reagents to their complementary sequences within the nucleic acid molecule of interest. Preferably, the

lengths of these nucleic acid reagents are at least 9 to 30 nucleotides. After incubation, all non-annealed nucleic acids are removed from the nucleic acid:fingerprint molecule hybrid. The presence of nucleic acids from the fingerprint tissue that have hybridized, if any such molecules exist, is then detected. Using such a detection scheme, the nucleic acid from the tissue or cell type of interest may be immobilized, for example, to a solid support such as a membrane, or a plastic surface such as that on a microtitre plate or polystyrene beads. In this case, after incubation, non-annealed, labeled nucleic acid reagents are easily removed. Detection of the remaining, annealed, labeled nucleic acid reagents is accomplished using standard techniques well-known to those in the art.

Alternative diagnostic methods for the detection of gene-specific nucleic acid molecules may involve their amplification, *e.g.*, by PCR (the experimental embodiment set forth in Mullis U.S. Patent No. 4,683,202 (1987)), ligase chain reaction (Barany, *Proc. Natl. Acad. Sci. USA*, 88:189-93 (1991)), self sustained sequence replication (Guatelli, *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:1874-78 (1990)), transcriptional amplification system (Kwoh, *et al.*, *Proc. Natl. Acad. Sci. USA*, 86:1173-77 (1989)), Q-Beta Replicase (Lizardi *et al.*, *Bio/Technology*, 6:1197 (1988)), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In one embodiment of such a detection scheme, a cDNA molecule is obtained from an RNA molecule of interest (*e.g.*, by reverse transcription of the RNA molecule into cDNA). Cell types or tissues from which such RNA may be isolated include any tissue in which wild-type fingerprint gene is known to be expressed, including, but not limited, to brain, cortex, subcortical region, cerebellum, brainstem, olfactory bulb, spinal cord, eye, Harderian gland, heart, lung, liver, pancreas, kidney, spleen, thymus, lymph nodes, bone marrow, skin, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, large intestine, cecum, testis, epididymis, seminal vesicle, coagulating gland, prostate gland, ovary, uterus and white fat. A sequence within the cDNA is then used as the template for a nucleic acid amplification reaction, such as a PCR amplification reaction, or the like. The nucleic

acid reagents used as synthesis initiation reagents (*e.g.*, primers) in the reverse transcription and nucleic acid amplification steps of this method may be chosen from among the gene nucleic acid reagents described herein. The preferred lengths of such nucleic acid reagents are at least 15-30 nucleotides. For detection of the amplified product, the nucleic acid amplification may be performed using radioactively or non-radioactively labeled nucleotides. Alternatively, enough amplified product may be made such that the product may be visualized by standard ethidium bromide staining or by utilizing any other suitable nucleic acid staining method.

Antibodies directed against wild-type or mutant gene peptides may also be used as disease diagnostics and prognostics. Such diagnostic methods, may be used to detect abnormalities in the level of CXCR6 protein expression, or abnormalities in the structure and/or tissue, cellular, or subcellular location of fingerprint CXCR6 protein. Structural differences may include, for example, differences in the size, electronegativity, or antigenicity of the mutant fingerprint CXCR6 protein relative to the normal fingerprint CXCR6 protein.

Protein from the tissue or cell type to be analyzed may easily be detected or isolated using techniques that are well known to those of skill in the art, including but not limited to western blot analysis. For a detailed explanation of methods for carrying out western blot analysis, see Sambrook, *et al.* (1989) *supra*, at Chapter 18. The protein detection and isolation methods employed herein may also be such as those described in Harlow and Lane, for example, (Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988)).

Preferred diagnostic methods for the detection of wild-type or mutant gene peptide molecules may involve, for example, immunoassays wherein fingerprint gene peptides are detected by their interaction with an anti-fingerprint gene-specific peptide antibody.

For example, antibodies, or fragments of antibodies useful in the present invention may be used to quantitatively or qualitatively detect the presence of wild-type or mutant gene peptides. This can be accomplished, for example, by immunofluorescence techniques employing a fluorescently labeled antibody (see below) coupled with light microscopic, flow cytometric, or fluorimetric detection. Such

techniques are especially preferred if the fingerprint gene peptides are expressed on the cell surface.

The antibodies (or fragments thereof) useful in the present invention may, additionally, be employed histologically, as in immunofluorescence or immunoelectron microscopy, for *in situ* detection of fingerprint gene peptides. *In situ* detection may be accomplished by removing a histological specimen from a patient, and applying thereto a labeled antibody of the present invention. The antibody (or fragment) is preferably applied by overlaying the labeled antibody (or fragment) onto a biological sample. Through the use of such a procedure, it is possible to determine not only the presence of the fingerprint gene peptides, but also their distribution in the examined tissue. Using the present invention, those of ordinary skill will readily perceive that any of a wide variety of histological methods (such as staining procedures) can be modified in order to achieve such *in situ* detection.

Immunoassays for wild-type, mutant, or expanded fingerprint gene peptides typically comprise incubating a biological sample, such as a biological fluid, a tissue extract, freshly harvested cells, or cells that have been incubated in tissue culture, in the presence of a detectably labeled antibody capable of identifying fingerprint gene peptides, and detecting the bound antibody by any of a number of techniques well known in the art.

The biological sample may be brought in contact with and immobilized onto a solid phase support or carrier such as nitrocellulose, or other solid support that is capable of immobilizing cells, cell particles or soluble proteins. The support may then be washed with suitable buffers followed by treatment with the detectably labeled gene-specific antibody. The solid phase support may then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on solid support may then be detected by conventional means.

The terms "solid phase support or carrier" are intended to encompass any support capable of binding an antigen or an antibody. Well-known supports or carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, gabbros, and magnetite. The nature of the carrier can be either soluble to some extent or insoluble for the purposes of the present invention.

The support material may have virtually any possible structural configuration so long as the coupled molecule is capable of binding to an antigen or antibody. Thus, the support configuration may be spherical, as in a bead, or cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface may be flat such as a sheet, test strip, etc. Preferred supports include polystyrene beads. Those skilled in the art will know many other suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

The binding activity of a given lot of anti-wild-type or -mutant fingerprint gene peptide antibody may be determined according to well known methods. Those skilled in the art will be able to determine operative and optimal assay conditions for each determination by employing routine experimentation.

One of the ways in which the gene peptide-specific antibody can be detectably labeled is by linking the same to an enzyme and using it in an enzyme immunoassay (EIA) (Voller, *Ric Clin Lab*, 8:289-98 (1978) ["The Enzyme Linked Immunosorbent Assay (ELISA)", *Diagnostic Horizons* 2:1-7, 1978, Microbiological Associates Quarterly Publication, Walkersville, Md.]; Voller, *et al.*, *J. Clin. Pathol.*, 31:507-20 (1978); Butler, *Meth. Enzymol.*, 73:482-523 (1981); Maggio (ed.), *Enzyme Immunoassay*, CRC Press, Boca Raton, Fla. (1980); Ishikawa, *et al.*, (eds.) *Enzyme Immunoassay*, Igaku-Shoin, Tokyo (1981)). The enzyme that is bound to the antibody will react with an appropriate substrate, preferably a chromogenic substrate, in such a manner as to produce a chemical moiety that can be detected, for example, by spectrophotometric, fluorimetric or by visual means. Enzymes that can be used to detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods that employ a chromogenic substrate for the enzyme. Detection may also be accomplished by visual comparison of the extent of enzymatic reaction of a substrate in comparison with similarly prepared standards.

Detection may also be accomplished using any of a variety of other immunoassays. For example, by radioactively labeling the antibodies or antibody fragments, it is possible to detect fingerprint gene wild-type, mutant, or expanded peptides through the use of a radioimmunoassay (RIA) (*see, e.g.*, Weintraub, B., Principles of Radioimmunoassays, Seventh Training Course on Radioligand Assay Techniques, The Endocrine Society, March, 1986). The radioactive isotope can be detected by such means as the use of a gamma counter or a scintillation counter or by autoradiography.

It is also possible to label the antibody with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wave length, its presence can then be detected due to fluorescence. Among the most commonly used fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine.

The antibody can also be detectably labeled using fluorescence emitting metals such as ^{152}Eu , or others of the lanthanide series. These metals can be attached to the antibody using such metal chelating groups as diethylenetriaminepentaacetic acid (DTPA) or ethylenediamine-tetraacetic acid (EDTA).

The antibody also can be detectably labeled by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction. Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, theromatic acridinium ester, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound may be used to label the antibody of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase and aequorin.

Throughout this application, various publications, patents and published patent applications are referred to by an identifying citation. The disclosures of these

publications, patents and published patent specifications referenced in this application are hereby incorporated by reference into the present disclosure to more fully describe the state of the art to which this invention pertains.

The following examples are intended only to illustrate the present invention and should in no way be construed as limiting the subject invention.

Examples

Example 1: Generation of Mice Comprising CXCR6 Gene Disruptions

To investigate the role of CXCR6, disruptions in CXCR6 genes were produced by homologous recombination. Specifically, transgenic mice comprising disruptions in CXCR6 genes were created. More particularly, as shown in Figure 4, a CXCR6-specific targeting construct based upon SEQ ID NO:1 or the sequence identified in GenBank as Accession No.: AF301018; GI:11342661, was created using as the targeting arms (homologous sequences) in the construct the oligonucleotide sequences identified herein as SEQ ID NO:3 or SEQ ID NO:4.

The targeting construct was introduced into ES cells derived from the 129/OlaHsd mouse substrain to generate chimeric mice. The F1 mice were generated by breeding with C57BL/6 females, and the F2 homozygous mutant mice were produced by intercrossing F1 heterozygous males and females.

Example 2: Expression Analysis

RT-PCR Expression. Total RNA was isolated from the organs or tissues from adult C57BL/6 wild-type mice. RNA was DNaseI treated, and reverse transcribed using random primers. The resulting cDNA was checked for the absence of genomic contamination using primers specific to non-transcribed genomic mouse DNA. cDNAs were balanced for concentration using HPRT primers.

The highest levels of RNA transcripts were detectable in spleen and lymph nodes.

Lower levels of RNA transcripts were also detectable in: whole brain, spinal cord, eye, Harderian glands, heart, lung, liver, pancreas, kidney, thymus, bone marrow, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, cecum, colon, testis, epididymis, seminal vesicle, prostate gland, ovary, uterus and white fat.

No RNA transcripts were detectable in: cortex, subcortical region, cerebellum, brainstem, olfactory bulb, skin and coagulating gland.

LacZ Reporter Gene Expression. In general, tissues from 7-12 week old heterozygous mutant mice were analyzed for lacZ expression. Organs from heterozygous mutant mice were frozen, sectioned (10 μ m), stained and analyzed for lacZ expression using X-Gal as a substrate for beta-galactosidase, followed by a Nuclear Fast Red counterstaining.

In addition, for brain, wholemount staining was performed. The dissected brain was cut longitudinally, fixed and stained using X-Gal as the substrate for beta-galactosidase. The reaction was stopped by washing the brain in PBS and then fixed in PBS-buffered formaldehyde.

Wild-type control tissues were also stained for lacZ expression to reveal any background or signals due to endogenous beta-galactosidase activity. The following tissues can show staining in the wild-type control sections and are therefore not suitable for X-gal staining: small and large intestines, stomach, vas deferens and epididymis. It has been previously reported that these organs contain high levels of endogenous beta-galactosidase activity.

LacZ expression was detected in urinary bladder and skin. In urinary bladder, a few cells in the lamina propria showed moderate staining. A small number of epidermal cells also showed X-Gal staining.

LacZ expression was not detected in: brain, spinal cord, sciatic nerve, eyes, thymus, spleen, lymph nodes, bone marrow, aorta, heart, lung, liver, pancreas, kidney, thyroid gland, pituitary gland, adrenal glands, skeletal muscle, testis, prostate gland, ovary and uterus.

Example 3: Physical Examination

A complete physical examination was performed on each mouse. Mice were first observed in their home cages for a number of general characteristics including activity level, behavior toward siblings, posture, grooming, breathing pattern and sounds, and movement. General body condition and size were noted as well identifying characteristics including coat color, belly color, and eye color. Following a visual inspection of the mouse in the cage, the mouse was handled for a detailed, stepwise

examination. The head was examined first, including eyes, ears, and nose, noting any discharge, malformations, or other abnormalities. Lymph nodes and glands of the head and neck were palpated. Skin, hair coat, axial and appendicular skeleton, and abdomen were also examined. The limbs and torso were examined visually and palpated for masses, malformations or other abnormalities. The anogenital region was examined for discharges, staining of hair, or other changes. If the mouse defecates during the examination, the feces were assessed for color and consistency. Abnormal behavior, movement, or physical changes may indicate abnormalities in general health, growth, metabolism, motor reflexes, sensory systems, or development of the central nervous system.

Example 4: Necropsy

Necropsy was performed on mice following deep general anesthesia, cardiac puncture for terminal blood collection, and euthanasia. Body lengths and body weights were recorded for each mouse. The necropsy included detailed examination of the whole mouse, the skinned carcass, skeleton, and all major organ systems. Lesions in organs and tissues were noted during the examination. Designated organs, from which extraneous fat and connective tissue have been removed, were weighed on a balance, and the weights were recorded. Weights were obtained for the following organs: heart, liver, spleen, thymus, kidneys, and testes/epididymides.

Example 5: Histopathological Analysis

Harvested organs were fixed in about 10% neutral buffered formalin for a minimum of about 48 hours at room temperature. Tissues were trimmed and samples taken to include the major features of each organ. If any abnormalities were noted at necropsy or at the time of tissue trimming, additional sample(s), if necessary, were taken to include the abnormalities so that it is available for microscopic analysis. Tissues were placed together, according to predetermined groupings, in tissue processing cassettes. All bones (and any calcified tissues) were decalcified with a formic acid or EDTA-based solution prior to trimming.

The infiltration of the tissues by paraffin was performed using an automated tissue processor. Steps in the cycle included dehydration through a graded series of ethanols, clearing using xylene or xylene substitute and infiltration with paraffin. Tissues were

embedded in paraffin blocks with a standard orientation of specified tissues within each block. Sections were cut from each block at a thickness of about 3-5 μ m and mounted onto glass slides. After drying, the slides were stained with hematoxylin and eosin (H&E) and a glass coverslip was mounted over the sections for examination.

5 **Example 6.0: Behavioral Analysis – Rotarod Test**

The Accelerating Rotarod was used to screen for motor coordination, balance and ataxia phenotypes. Mice were allowed to move about on their wire-cage top for 30 seconds prior to testing to ensure awareness. Mice were placed on the stationary rod, facing away from the experimenter. The "speed profile" programs the rotarod to reach 60 rpm after six minutes. A photobeam was broken when the animal fell, which stopped the test clock for that chamber. The animals were tested over three trials with a 20-minute rest period between trials, after which the mice were returned to fresh cages. The data was analyzed to determine the average speed of the rotating rod at the fall time over the three trials. A decrease in the speed of the rotating rod at the time of fall compared to wild-types indicated decreased motor coordination possibly due to a motor neuron or inner ear disorder.

Example 6.1: Behavioral Analysis – Startle Test

The startle test screens for changes in the basic fundamental nervous system or muscle-related functions. This includes changes in 1) hearing - auditory processing; 2) sensory and motor processing - related to the auditory circuit and culminating in a motor related output; and 3) motor abnormalities, including skeletal muscle or motor neuron related changes. The startle reflex is a short-latency response of the skeletal musculature elicited by a sudden auditory stimulus. The startle reflex is seen across many species, making the startle response test a useful animal model for studying abnormalities in the neural control of simple behaviors and searching for treatments and causes of those abnormalities. In rats or mice, the response is usually measured in a response chamber, which allows the measurement of the whole-body flinch elicited by the stimulus. Similar stimuli are used to test the response in humans, where a blink response is measured using electromyography of the orbicularis oculi muscle.

One component of the startle reflex test is prepulse inhibition (PPI). PPI is the reduction or gating of the startle reflex response produced by a weak prestimulus

presented at a brief interval, usually between 30-500ms, before the startle eliciting stimulus. Both rats and humans exhibit a graded increase in PPI with increasing prepulse intensities.

Deficits in PPI are observed in human schizophrenic patients. Deficits in PPI have been associated with dopamine overactivity, as shown by the ability to produce a loss of PPI in rats treated with dopamine agonists, such as apomorphine. PPI can be restored in apomorphine treated rats by antipsychotics in a manner that correlates with clinical antipsychotic potency and D₂ receptor affinity. It is also believed that neural modulation of PPI in rats is affected by circuitry linking the hippocampus (HPC), the nucleus accumbens (NAC), the subpallidum, and the pontine reticular formation. Aside from dopaminergic involvement in PPI and sensory gating, both forebrain glutamatergic and serotonergic systems have been implicated in the pathophysiology of schizophrenia and the action of atypical antipsychotics, and both glutamatergic and serotonergic activity are important substrates modulating PPI in rats. Non competitive NMDA glutamate receptor antagonists and serotonin receptor (particularly 5-HT_{1B}) agonists have both been shown to reduce PPI in rats.

Genetic factors may be critical determinants of sensorimotor gating in rats. This has been supported by studies showing strain related differences in the dopaminergic modulation of PPI, as well as the production through inbreeding of strains of rats whose behavior was either apomorphine-sensitive or insensitive. Rats having a disruption of the 5-HT_{1B} were reported to have slightly elevated basal PPI compared to wild-type controls, indicating a tonic regulation of PPI by 5-HT_{1B}. This conclusion was supported by research showing that a 5-HT_{1A/1B} agonist reduced PPI in wild-type mice, but not in the 5-HT_{1B} knockouts. The investigation of the effects on PPI of disruptions of other genes could be a valuable tool for understanding the role of particular gene products in the regulation of PPI and sensorimotor gating.

The connection between the abnormalities in sensorimotor gating in schizophrenic patients and PPI are supported by the belief that brain regions frequently implicated in the pathophysiology of the disorder are also involved in the regulation of PPI. Abnormalities at several levels of the startle gating circuitry, including the

hippocampus, nucleus accumbens, striatum, globus pallidus, and thalamus, have been noted in schizophrenic patients.

The mice were tested as follows:

Sound Response Profile. The mice were tested in a San Diego Instruments SR-

5 LAB sound response chamber. Each mouse was exposed to 9 stimulus types that were repeated in pseudo-random order ten times during the course of the entire 25 minute test. The stimulus types in decibels were: p80, p90, p100, p110, p120, pp80, p120, pp90, p120, pp100, and p120; where p=40 msec pulse, pp=20 msec prepulse. The length of time between a prepulse and a pulse was 100 msec (onset to onset). The mean Vmax of
10 the ten repetitions for each trial type was computed for each mouse.

Pre-Pulse Inhibition. The % prepulse inhibition compared to p120 alone was computed for each mouse at three prepulse levels from the mean Vmax values. This was computed by determining the mean “p120”, “pp80p120”, “pp90p120”, and “pp100p120” value for each mouse and then producing the ratios of % inhibition.

15 **Example 6.2: Behavioral Analysis - Hot Plate Test**

The hot plate analgesia test was designed to indicate an animal's sensitivity to a painful stimulus. The mice were placed on a hot plate of about 55.5°C, one at a time, and latency of the mice to pick up and lick or fan a hindpaw was recorded. A built-in timer was started as soon as the subjects were placed on the hot plate surface. The timer was
20 stopped the instant the animal lifted its paw from the plate, reacting to the discomfort. Animal reaction time was a measurement of the animal's resistance to pain. The time points to hindpaw licking or fanning, up to a maximum of about 60-seconds, was recorded. Once the behavior was observed, the animal was immediately removed from the hot plate to prevent discomfort or injury.

25 **Example 6.3: Behavioral Analysis – Tail Flick Test**

The tail-flick test is a test of acute nociception in which a high-intensity thermal stimulus is directed to the tail of the mouse. The time from onset of stimulation to a rapid flick/withdrawal from the heat source is recorded. This test produces a simple nociceptive reflex response that is an involuntary spinally mediated flexion reflex.

30 **Example 6.4: Behavioral Analysis – Open Field Test**

The Open Field Test was used to examine overall locomotion and anxiety levels in mice. Increases or decreases in total distance traveled over the test time are an indication of hyperactivity or hypoactivity, respectively.

5 The open field provides a novel environment that creates an approach-avoidance conflict situation in which the animal desires to explore, yet instinctively seeks to protect itself. The chamber is lighted in the center and has no places to hide other than the corners. A normal mouse typically spends more time in the corners and around the periphery than it does in the center. Normal mice however, will venture into the central regions as they explore the chamber. Anxious mice spend most of their time in the
10 corners, with almost no exploration of the center, whereas bold mice travel more, and show less preference for the periphery versus the central regions of the chamber.

Each mouse was placed gently in the center of its assigned chamber. Tests were conducted for 10 minutes, with the experimenter out of the animals' sight. Immediately following the test session, the fecal boli were counted for each subject: increased boli are
15 also an indication of anxiety. Activity of individual mice was recorded for the 10-minute test session and monitored by photobeam breaks in the x-, y- and z-axes. Measurements taken included total distance traveled, percent of session time spent in the central region of the test apparatus, and average velocity during the ambulatory episodes. Increases or decreases in total distance traveled over the test time indicate hyperactivity or
20 hypoactivity, respectively. Alterations in the regional distribution of movement indicates anxiety phenotypes, i.e., increased anxiety if there is a decrease in the time spent in the central region.

Example 6.5: Behavioral Analysis – Metrazol Test

To screen for phenotypes involving changes in seizure susceptibility, the Metrazol
25 Test was used. About 5mg/ml of Metrazol was infused through the tail vein of the mouse at a constant rate of about 0.375ml/min. The infusion caused all mice to experience seizures. Those mice who entered the seizure stage the quickest were thought to be more prone to seizures in general.

The Metrazol test can also be used to screen for phenotypes related to epilepsy.
30 Seven to ten adult wild-type and homozygote males were used. A fresh solution of about 5mg/ml pentylenetetrazole in approximately 0.9% NaCl was prepared prior to testing.

Mice were weighed and loosely held in a restrainer. After exposure to a heat lamp to dilate the tail vein, mice were continuously infused with the pentylenetetrazole solution using a syringe pump set at a constant flow rate. The following stages were recorded: first twitch (sometimes accompanied by a squeak), beginning of the tonic/clonic seizure, tonic extension and survival time. The dose required for each phase was determined and the latency to each phase was determined between genotypes. Alterations in any stage may indicate an overall imbalance in excitatory or inhibitory neurotransmitter levels.

Example 6.6: Behavioral Analysis – Tail Suspension Test

The tail suspension test is a single-trial test that measures a mouse's propensity towards depression. This method for testing antidepressants in mice was reported by Steru *et al.*, (1985, *Psychopharmacology* 85(3):367-370) and is widely used as a test for a range of compounds including SSRI's, benzodiazepines, typical and atypical antipsychotics. It is believed that a depressive state can be elicited in laboratory animals by continuously subjecting them to aversive situations over which they have no control. It is reported that a condition of "learned helplessness" is eventually reached.

Mice were suspended on a metal hanger by the tail in an acoustically and visually isolated setting. Total immobility time during the six-minute test period was determined using a computer algorithm based upon measuring the force exerted by the mouse on the metal hanger. An increase in immobility time for mutant mice compared to wild-type mice may indicate increased "depression." Animals that ceased struggling sooner may be more prone to depression. Studies have shown that the administration of antidepressants prior to testing increases the amount of time that animals struggle.

Example 7: Hematological Analysis

Blood samples were collected via a terminal cardiac puncture in a syringe. About one hundred microliters of each whole blood sample were transferred into tubes pre-filled with EDTA. Approximately 25 microliters of the blood was placed onto a glass slide to prepare a peripheral blood smear. The blood smears were later stained with Wright's Stain that differentially stained white blood cell nuclei, granules and cytoplasm, and allowed the identification of different cell types. The slides were analyzed microscopically by counting and noting each cell type in a total of 100 white blood cells.

The percentage of each of the cell types counted was then calculated. Red blood cell morphology was also evaluated.

Microscopic examinations of blood smears were performed to provide accurate differential blood leukocyte counts. The leukocyte differential counts were provided as the percentage composition of each cell type in the blood.

Example 8: Serum Chemistry

Blood samples were collected from mice via a terminal cardiac puncture with a syringe. The blood sample was converted to serum by centrifugation in a serum tube with a gel separator. Each serum sample was then analyzed for the following analytes: alanine aminotransferase; albumin; alkaline phosphatase; bicarbonate; total bilirubin; blood urea nitrogen; calcium; chloride; cholesterol; creatinine kinase; creatinine; globulin; glucose; high density lipoproteins; lactate dehydrogenase; low density lipoproteins; osmolality; phosphorus; potassium; total protein; sodium; and triglycerides.

Non-terminal blood samples were collected via retro-orbital venous puncture in capillary tubes. This procedure yielded approximately 200 μ L of whole blood that is transferred into a serum tube with a gel separator for serum chemistry analysis.

Example 9: Densitometric Analysis

Mice were euthanized and analyzed using a PIXImusTM densitometer. An x-ray source exposed the mice to a beam of both high and low energy x-rays. The ratio of attenuation of the high and low energies allowed the separation of bone from soft tissue, and, from within the tissue samples, lean and fat. Densitometric data including Bone Mineral Density (BMD presented as g/cm²), Bone Mineral Content (BMC in g), bone and tissue area, total tissue mass, and fat as a percent of body soft tissue (presented as fat %) were obtained and recorded.

Example 10: Embryonic Development

Animals are genotyped using one of two methods. The first method uses the polymerase chain reaction (PCR) with target-specific and Neo primers to amplify DNA from the targeted gene. The second method uses PCR and Neo primers to “count” the number of Neo genes present per genome.

If homozygous mutant mice are not identified at weaning (3-4 weeks old), animals were assessed for lethality linked with the introduced mutation. This evaluation included embryonic, perinatal or juvenile death.

Newborn mice were genotyped 24-48 hours after birth and monitored closely for any signs of stress. Dead/dying pups were recorded and grossly inspected and if possible, genotyped. In the case of perinatal death, late gestation embryos (~E19.5, i.e., 19.5 days post-coitum) or newborn pups were analyzed, genotyped and subject to further characterization.

If there was no evidence of perinatal or juvenile lethality, heterozygous mutant mice were set up for timed pregnancies. Routinely, E10.5 embryos are analyzed for gross abnormalities and genotyped. Depending on these findings, earlier (routinely >E8.5) or later embryonic stages are characterized to identify the approximate time of death. If no homozygous mutant progeny are detected, blastocysts (E3.5) are isolated, genotyped directly or grown for 6 days in culture and then genotyped. Any suspected genotype-related gross abnormalities are recorded.

Example 11: Fertility

The reproductive traits of male and female homozygous mutant mice are tested to identify potential defects in spermatogenesis, oogenesis, maternal ability to support pre- or post-embryonic development, or mammary gland defects and ability of the female knockout mice to nurse their pups.

Homozygous mutant (-/-) mice of each gender were set up in a fertility mating with either a wild-type (+/+) mate or a homozygous mutant mouse of the opposite gender at about seven to about ten weeks of age. The numbers of pups born from one to three litters were recorded at birth. Three weeks later, the live pups were counted and weaned.

Males and females were separated after they had produced two litters or at six months (26 weeks) of age, whichever comes first.

Example 12: Cytofluorometric Analyses

Cells from thymus, lymph nodes, and spleen tissues were isolated from wild type and mutant mice and dispersed into a single cell suspension. The red blood cells were removed by lysis with Tris/NH₄Cl solution for 5 minutes at room temperature. The cell suspension was filtered with a nylon mesh and washed twice with a staining medium,

which was HBSS with reduced phenol red, sodium azide, BSA, and EDTA. 0.5×10^6 cells/25 μ l/staining were incubated with 1 μ g/10 μ l/staining of PE- or FITC-labeled antibodies (PharMingen, San Diego, Calif.) for 15 minutes on ice, washed once and fixed with 0.5% formamide in staining medium. Cytometric analyses were performed using
5 FACSscan (Becton Dickinson) as described previously (Hanna Z et al., *Mol. Cell. Biol.*, 1994, 14:1084-1094). About 20,000 cells were recorded in each staining.

Example 13: Metabolic Screens

Female mice of about 8 weeks old were put on a high fat diet (about 42% calories, Adjusted Calories Diet #88137, Harlan Teklad, Madison, WI) and subjected to a Glucose
10 Tolerance Test about 8 weeks later and Densitometric measurements about 10 weeks later. The body weights and lengths (metrics) were also recorded during the course of high fat diet challenge.

Glucose Tolerance Test (GTT): Mice were fasted for about 3 hours and tail vein blood glucose levels were measured before injection by collecting about 5 to 10
15 microliters of blood from the tail tip and using glucometers (Glucometer Elite, Bayer Corporation, Mishawaka, IN). The glucose values were used for time $t=0$. Mice were weighed at $t=0$ and glucose was administered orally or by intra-peritoneal injection at a dose of about 2 grams per kilogram of body weight. Plasma glucose concentrations were measured at about 15, 30, 60, 90, and 120 minutes after injection by the same
20 method used to measure basal ($t=0$) blood glucose.

The glucose levels may represent the ability of the mouse to secrete insulin in response to an elevated plasma glucose concentration or the ability of certain tissues, such as, for example, muscle, liver and adipose tissues, to uptake glucose.

Densitometric Analysis: Mice were anaesthetized with isoflurane and analyzed
25 using a PIXImusTM densitometer. An x-ray source exposed the mice to a beam of both high and low energy x-rays. The ratio of attenuation of the high and low energies allowed the separation of bone from soft tissue, and, from within the tissue samples, lean and fat. Densitometric data including Bone Mineral Density (BMD presented as g/cm^2), Bone Mineral Content (BMC in g), bone and tissue area, total tissue mass, and fat as a
30 percent of body soft tissue (presented as fat %) were obtained and recorded.

Metrics: Body lengths and body weights were recorded before and during the high fat diet challenge.

For all the data collected, two-tailed pair-wise statistical significance was established using a Student t-test. Statistical significance is defined as $P = 0.05$.

5 **Example 14: Pain and Nociception**

Formalin Test. The Formalin test for nociception involves injecting a noxious substance, about 3% Formalin solution, into the plantar surface of the mouse's hindpaw. The mouse reacts to the Formalin injection (by licking and flinching the injected hindpaw, for example). An automated system is used to detect the number of times the mouse flinches over a period of about one hour. The response to Formalin injection occurs as two distinct phases. Phase one occurs within about the first 10 minutes of the test and is thought to be the result of C-fiber activation due to the chemical stimulation of the nociceptors. Phase two occurs within about 11-60 minutes following the injection. Phase two appears to be due to a neurogenic inflammatory reaction within the injected paw and functional changes in the dorsal horn of the spinal cord.

Homozygous mutant (-/-) or heterozygous (-/+) mice showing a difference in the response to Formalin, relative to wild-type control mice may indicate a role of CXCR6 in nociception.

Paw Thermal Test. Nociception in the paw thermal test is accomplished by administering heat generated from a radiant bulb. About 12.5 μL of Complete Freund's Adjuvant (CFA) solution is injected into the plantar surface of a paw. After about 24 hours, mice are placed into test chambers and allowed to acclimate to the chamber for a minimum of about 30 minutes, or until exploratory and grooming behavior cease. A radiant bulb is positioned under a hind paw of the mouse, such that a focused light beam contacts the hind paw and delivers a heat stimulus. The mouse is observed for a response of either a stomp action or a sharp withdrawal of the paw. An automatic motion sensor stops the heat stimulus when the mouse responds. The response latency is recorded.

Homozygous mutant (-/-) or heterozygous (-/+) mice exhibiting a significant difference in response latency may indicate a role of CXCR6 in pain perception.

Mechanical Sensitivity Test. The nociception stimulus in the mechanical sensitivity test is the force of a filament applied to the plantar surface of both hind paws.

About 12.5 μ L of Complete Freund's Adjuvant (CFA) solution is injected into the plantar surface of a paw. After approximately 28 hours, mice are placed into test chambers and allowed to acclimate to the chamber for a minimum of about 30 minutes, or until exploratory and grooming behavior cease. A filament is then brought into contact with the paw. The filament touches the plantar surface of the hind paws and begins to exert an upward force below the threshold of feeling. The force increased at a rate of about 0.25 grams per second until the mouse removes his hindpaw or until the maximum force of about 5.0 grams is reached in approximately 20 seconds. The latency for the mouse to remove the hindpaw is recorded.

Neuropathic Pain Test. To investigate the effect of the CXCR6 disruption in the development of neuropathic pain, groups of about 12 male mice are tested.

Under normal conditions, each mouse is tested for its mechanosensory (tactile) response using the calibrated von Frey hairs (filament) test and its thermal sensitivity using the Hargreaves test (see Hargreaves *et al.*, 1988, *Pain* 32:77-88) on days -1 and 0 before nerve injury. Mechanical pain tests are conducted first, followed by thermal pain tests, with all data recorded. Neuropathic pain is then induced by either spinal nerve ligation per the Chung model (see Kim and Chung, 1992, *Pain* 50(3):355-363) or sciatic nerve injury (*i.e.*, chronic constriction injury). On about days 2, 4, 6, 8, 10 and 12, each mouse is subjected to two pain behavioral tests, with all data recorded.

On about day 12, mice are given about 100 mg/kg of gabapentin via intraperitoneal injection. About 60 to about 90 minutes post injection, mice are subjected to the two pain behavioral tests, with all data recorded.

Mice are then necropsied. Certain tissues are immediately dissected, including the brain (mainly the thalamus), spinal cord and dorsal root ganglia. Tissues are preserved in RNA Later Solution and frozen at -80 degrees Celsius, for later analysis. Homozygous mutant (-/-) or heterozygous (-/+) mice that exhibit differences, as compared to wild-type controls, in response latencies or that exhibit differences in response to pain testing after gabapentin administration, may indicate that CXCR6 plays a role in neuropathic pain perception. Mice having a disruption in the CXCR6 gene, according to the present invention may be used to screen for nociceptive agents and known compounds useful for treating pain.

Example 15: Cutaneous Allergy (aka Contact Dermatitis) Assay

Cutaneous contact allergy can be broadly categorized as allergic contact dermatitis (ACD), requiring multiple exposures to the allergen, and irritant contact dermatitis (ICD), which occurs following an initial exposure. Both ACD and ICD involve a local inflammatory response mediated by leukocytes such as neutrophils, monocytes, macrophages and mast cells. Unlike ICD, ACD also involves a T-cell component, since the affected individual must be exposed to the allergen more than one time. These inflammatory reactions involve a complex interplay between blood borne cells, blood vessels and the affected tissue itself.

Allergic Contact Dermatitis (ACD) Assay Procedure: On day 0, wild-type and homozygous mutant mice have their abdomen shaved and are primed with the allergen, a 1% (weight/volume) solution of fluorescein isothiocyanate (FITC; in a 1:1 solution of acetone and dibutylphthalate). Priming is performed by placing 400 microliters of the FITC solution on the shaved abdomen and 10 microliters of the solution on each footpad. On day 6, the ear thickness of each mouse is measured with an engineer's micrometer to establish a baseline ear thickness. To each mouse, 10 microliters of the 1% FITC solution is placed epicutaneously on one ear. The acetone:dibutylphthalate solvent (without FITC) is added to the other ear as a control. Ear swelling is estimated by determining the change in ear thickness from baseline to 24 hours post-challenge. Ear swelling is indicative of an allergic response to the antigen. Histological changes, including changes in the cell types infiltrating the ear, may also be evaluated to determine changes in the type of inflammation. Other means of measuring inflammation, such as thermogenic imaging to gauge temperature changes, may also be used.

Irritant Contact Dermatitis (ICD) Assay Procedure: On day 0, the ear thickness of each mouse (wild-type and homozygous mutant) is measured with an engineer's micrometer. Ten microliters of a 2% croton oil (volume/volume) solution (in a 4:1 acetone:olive oil solvent) is applied epicutaneously to one ear of the mice. The solvent (without croton oil) is applied on the other ear as a control. After 24 hours, the ear thickness of each mouse is measured (see ACD procedure above).

The CXCR6 gene may have an immunologically related phenotype and, specifically, may be involved in cutaneous allergy response, and that compounds targeted to the CXCR6 gene or gene product may be effective in modulating contact allergy response or symptoms.

5

As is apparent to one of skill in the art, various modifications of the above embodiments can be made without departing from the spirit and scope of this invention. These modifications and variations are within the scope of this invention.